



Potential of wild plant Artemisia judaica L. as sustainable source of antioxidant and antimicrobial compounds

Yasser A. El-Amier^{1*}, Abd El-Nasser S. Al Borki², Shrouk A. Elagami¹

¹Botany Department, Faculty of Science, Mansoura University, 35516, Mansoura, Egypt, ²Botany Department, Faculty of Science and Arts, Benghazi University, Agdabia, Libya

ABSTRACT

Artemisia judaica L. (Shih Balady, family Asteraceae), is a medicinal plant that grows in the desert, commonly used as tea by population in Egypt Sinai and of Saudi Arabia. The present study aims to evaluate antimicrobial and antioxidant activities of aerial parts of A. judaica collected from the inland desert (Wadi Hagoul) of Egypt. Total phenolics, tannins, alkaloids, flavonoids, and saponins were measured in the plant of interest. According to the reduction of DPPH the anti-oxidant activity was measured. Different extracts of A. judaica were screened for their antimicrobial activity against nine pathogenic microorganisms using filter paper disc assay. The secondary metabolites of A. judaica were examined and proved that it was rich in tannins and phenolics (13.29 and 7.62 mg/g dw). The IC $_{50}$ values of A. judaica extract was 1.78 mg.ml⁻¹ compared to standard catechol (0.15 mg.ml⁻¹). In the present study, the extracts of A. judaica showed unusual activity depending on the type of bacteria and fungi. Also, ethyl acetate and methyl alcohol extracts showed considerable broad spectrum (77.78%, each) against both bacteria and fungi, followed by acetone, then methylene chloride and petroleum ether extracts. This study showed that A. judaica extracts could be used as antimicrobial substances in pharmaceutical and food preservation systems.

Received: January 30, 2019 Accepted: March 18, 2019 Published: March 21, 2019

*Corresponding Author: Dr. Yasser Ahmed El-Amier Email: yasran@mans.edu.eg

KEYWORDS: Artemisia judaica, asteraceae antimicrobial, antioxidant, secondary compounds

INTRODUCTION

Nearly all cultures in the world depends on medicinal plants medicinal purpose. Around 60% of population continue to use herbal extraction in health care [1,2]. Many species of the genus Artemisia have been identified as aromatic and used in medicine [3]. Artemisia Judaica L. belongs to the family Asteraceae (Compositae) and known as wormwood (Arabic name, Shih Balady). It is a fragranced shrub that widely grows in the Arabian area [4]. A. judaica is used as a tea commonly by population in Egypt Sinai and of Saudi Arabia. In the Arabic area traditional medicine; A. judaica is used as a treatment for cardiovascular health, and many other diseases and dysfunctions [5, 6].

Secondary plant metabolites are active chemical defense against pathogens as well as they play an essential role in plant defense against herbivory or other interspecies defenses [7-10]. Phytochemical analysis of A. *judaica* which carried out by many authors [11-14] showed antioxidant activity [15,16], anti-malarial,

antibacterial, anti-inflammatory [17,18], biopesticide [19]. In the current paper, we investigate the potential antioxidant and antimicrobial properties of A. *judaica* collected from the inland desert (Wadi Hagoul) of Egypt, to evaluate their medicinal potentiality and their future industrial uses.

MATERIALS AND METHODS

Plant Material

A. *judaica* aerial parts were collected from populations which grow in Eastern desert, wadi Hagul, Egypt during the flowering period (March 2018). The plant species were identified according to Boulos [4].

Phytochemical Analysis

A. *judaica* was phytochemically tested for secondary plant metabolites, tannins, alkaloids, flavonoids, saponins and total phenolic compounds [20-24].

Copyright: © The authors. This article is open access and licensed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.o/) which permits unrestricted, use, distribution and reproduction in any medium, or format for any purpose, even commercially provided the work is properly cited. Attribution — You must give appropriate credit, provide a link to the license, and indicate if changes were made.

Determination of the Radical Scavenging Activity

The antioxidant activity by DPPH was determined as described by Miguel [25].

Antimicrobial Activity Assessment

Preparation of the crude extracts

Plant extract was prepared by standard methods of Freedman *et al.* [26] and Su and Horvat [27]. In each solvent, the treated filtrate evaporates, and the dried residue was dissolved in dimethyl sulfoxide (DMSO) and reserved at -20°C for future use [28].

Antibacterial Assay

The plant extracts were examined for the presence of antimicrobial bioactivity by the method of Cappuccino and Sherman [29] using different bacterial species (Bacillus subtilis, Escherichia coli, Klebsiella pneumonia, Staphylococcus aureus, and Streptococcus pyogenes) and fungi (Candida albicans, Aspergillus niger, Aspergillus fumigatus, and Mucor spp.). From the Laboratory of Bacteriology, Department of Botany, Faculty of Science, Mansoura University, Egypt the tested samples were taken.

The filter paper discs are prepared with a diameter of 5 mm and sterilized in the autoclave for 20-30 minutes. Then taken by sterile forceps, soaked in DMSO solution and then placed over the surface of the inoculated nutrient agar. In 37°C the Petri dishes were incubated for 24 hours, While in fungi, A sterile disc set over the surface of the inoculated potato dextrose agar and incubated for 3-4 days at 28°C. After incubation for period of time, the inhibition zones diameter (mm) was calculated to record the clear zone to determine the most active extract against pathogenic microorganisms.

RESULTS AND DISCUSSION

Phytochemical Constituents

Plants are the basis of many natural products such as; alkaloids, amines, cyanogenic glycosides, flavonoids, glucosinolates, phenylpropanes, terpenes, saponins, steroids, etc. It is estimated that there are more than 200,000 chemical structures is synthesized by plants [30,31]. Analysis of secondary metabolites of A. Judaica showed high values of tannins and phenolics (13.29 and 7.62 mg/g dry weight, respectively). Besides, saponins, alkaloids, and flavonoids (3.55, 2.84 and 1.63 mg/g dry weight, respectively) showed comparable contents, (Figure 1). These results were consistent (compatible) with the results of Essiett and Akpan [32] on Aspilia africana and Tithonia diversifolia (Asteraceae) and Erol-Dayi et al. [33] on Centaurea species, but they were lower than some wild plants such as Anthemis arvensis, Artemisia campestris, Senecio glaucus and Urospermum picroides [34-36]. It is not surprising to discover that secondary metabolites of the plant have medicinal activities, as most of the prescriptions have plant based origin [30].

Antioxidant Activity

Most organisms own antioxidant systems against ROS [37]. Excessive ROS states have main action in diseases like idiopathic pulmonary fibrosis and respiratory distress syndrome [38]. The evaluation of the antioxidant activity evaluation in plant extracts is shown in Figure 2. By increasing the concentration of plant extract, a continuous corresponding increase in the scavenging activity. In methanol extract and standard antioxidant, the scavenging activities were 55.43% and 82.72%, respectively at 4000 µg.ml⁻¹.

The IC₅₀ values of A. *judaica* extract was 1.78 mg.ml⁻¹ compared to standard catechol (0.15 mg.ml⁻¹). According to Al-Ismail *et al.* [39], A. *judaica* extract showed moderate scavenging activity (IC₅₀ \geq 1 and \leq 2 mg/ml). These results suggest that methanol extract of A. *judaica* has an obvious effect on scavenging of DPPH radical. Similar results were reported by Pandey and Singh [40] on the same genus Artemisia, El-Amier and Abdullah [41] on Calligonum comosum and Salem *et al.* [42] on Silybum marianum.

The plant extract potent antioxidant activity can be mainly attributed to phenolic content, as a result of heir hydroxyl group, and/or flavonoids that react with DPPH radical by donating hydrogen atom the free radicals [43], while a highly

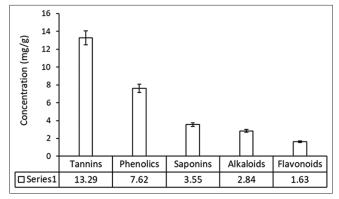


Figure 1: The concentration of the active constituents in mg/g dry weight for the *Artimisa judaica*

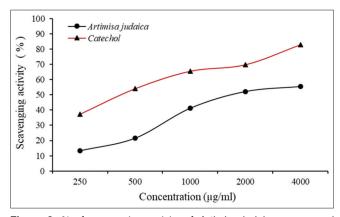


Figure 2: % of scavenging activity of Artimisa judaica extract and natural antioxidant catechol

J Exp Sci • 2019 • Vol 10

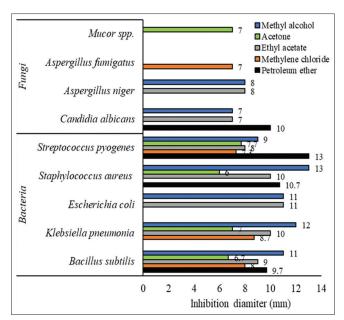


Figure 3: Antimicrobial activity of a different extract of *Artimisa judaica* against tested pathogen

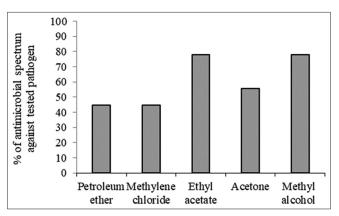


Figure 4: Percent of the antimicrobial spectrum of *Artimisa judaica* extracts against tested pathogen

positive correlation between total phenolic content and antioxidant activity was established in the case of many plant species [11,44,45].

Antimicrobial Activity Assessment

The plants showed well defined action agains microbes [46, 47]. In this investigation, the petroleum ether extract showed higher activity (Figure 3). The *Stroptococcus pyogenes* was the most potent inhibitor followed by *Staphylococcus aureus* and *Bacillus subtilis*. Methylene chloride does not act on *Escherichia coli* and *S. aureus* but inhibiting others. The ethyl acetate and methyl alcohol extracts inhibit all pathogenic bacteria with different rates. Previous reports supports these results [48, 49]. These results were similar to those of *Ocimum sanctum Xanthoxylum armatum*, *Cinnamomum zeylanicum*, and *Origanum majorana* as investigated by Joshi, et al. [50] and Iranbakhsh et al. [51] in *Datura stramonium* (B. subtilis, E. coli, and S. aureus).

On the other hand, petroleum ether extract had no antifungal activities on all the pathogenic fungi except *Candidia albicans* (Figure 3). The methylene chloride extract inhibited the growth of *Aspergillus fumigatus* only, and ethyl acetate extract does not affect both *A. fumigatus* and *Mucor* spp. But hindered others. No antifungal activities were detected with acetone extract on all the pathogenic fungi except *Mucor* spp. Methyl alcohol extract inhibited the growth of *C. albicans* and *A. fumigatus* (Figure 3). Abdel-Sattar *et al.* [52] reported *Achillea biebersteinii* and *Vernonia schimperii* was active against the same tested fungi strain. Also, these results were compared with other investigators [53-55].

The extracts of A. *judaica* showed action (77.78%, each) against microbes (Figure 4), followed by acetone (55.56%) then methylene chloride and petroleum ether extracts (44.44%, each). The pathogen *S. aureus* and *E. coli* (13 and 11 mm, respectively) were the sensitive bacteria in ethyl acetate and methylene chloride extracts, respectively, whereas *C. albicans* was sensitive fungi in petroleum ether extract (10 mm).

There are many actions for flavonoids in medicinal field [56]. Tannins are capable of inhibiting the digestive enzymes activity, and the nutritional effects of tannins are mainly linked with their interaction with protein [57]. Tannin-protein complexes are insoluble, and the protein digestibility is decreased [58]. Plant saponins are well known for their medicinal actions [59]. Alkaloids have profound effects on diseases [60, 61].

CONCLUSION

In the present study, A. judaica phytochemical analysis showed increase in the secondary compounds, and their extract showed moderate scavenging activity. The petroleum ether extract of A. judaica exhibited pronounced activity against bacteria than fungi. The methyl alcohol and ethyl acetate extracts showed inhibitory action on bacteria and fungi. The pathogen S. pyogenes was the most potent inhibitor followed by S. aureus and B. subtilis. Also, the pathogen S. aureus, E. coli and C. albicans were the most sensitive microorganism. This study revealed that A. judaica extracts could be an alternative in pharmaceutical and food preservation systems.

CONTRIBUTION OF AUTHORS

All Authors contribute in the study equally either in collection and analyzing of samples or preparation and writing the manuscript.

REFERENCES

- Abdel-Sattar EA, Harraz FM, Gayed SH. Antimicrobial activity of extracts of some plants collected from the Kingdom of Saudi Arabia. Journal of Medical Science. 2008:15(1): 25-33.
- Abd-Elhady HK, Insecticidal activity and chemical composition of essential oil from Artemisia judaica L. against Callosobruchus maculatus (F.) (CO-LEOPTERA: BRUCHIDAE). Journal of Plant Protection Research, 2012; 52(3): 347–352.
- Abu-Darwish MS, Cabral C, Gonçalves MJ, Cavaleiro C, Cruz MT, Zulfiqar A, Khan IA, Efferth T, Salgueiro L. Chemical composition and

- biological activities of *Artemisia judaica* essential oil from southern desert of Jordan. Journal of Ethnopharmacology. 2016; 191:161-8.
- Acheuk F, Lakhdari W, Abdellaoui K, Belaid M, Allouane R, Halouane F. Phytochemical study and bioinsecticidal effect of the crude ethonolic extract of the Algerian plant *Artemisia judaica* L.(Asteraceae) against the black bean aphid, Aphis fabae Scop. Poljoprivreda i Sumarstvo. 2017; 63(1): 95-104.
- 5. Alamgir AN. Therapeutic use of medicinal plants and their extracts: volume 1: pharmacognosy. 2017 Sep 6, Springer.
- Al-Ismail K, Herzallah SM, Rustom AS. Antioxidant activities of some edible wild Mediterranean plants. Italian Journal of Food Science. 2007; 19(3): 287-296.
- Allen RD, Webb RP, Schake SA. Use of transgenic plants to study antioxidant defenses. Free Radical Biology and Medicine. 1997; 23(3): 473-9.
- Boham BA, Kocipai-Abyazan R. Flavonoids and condensed tannins from leaves of Hawaiian Vaccinium vaticulatum and V. calycinium. Pacific Science. 1974; 48: 458-63.
- Boulos L. Flora of Egypt. Vol. III (Verbenaceae-Compositae). 2002, Al-Hadara Publishing, Cairo, Egypt.
- Calliste CA, Trouillas P, Allais DP, Simon A, Duroux JL. Free radical scavenging activities measured by electron spin resonance spectroscopy and B16 cell antiproliferative behaviors of seven plants. Journal of Agricultural and Food Chemistry. 2001; 49(7): 3321-7.
- 11. Cappuccino JG, Sherman N. Microbiology: a laboratory manual.
- Carnovale E, Lugaro E, Marconi E. Protein quality and antinutritional factors in wild and cultivated species of *Vigna* spp. Plant Foods for Human Nutrition. 1991; 41(1): 11-20.
- Chiang YM, Chuang DY, Wang SY, Kuo YH, Tsai PW, Shyur LF. Metabolite profiling and chemopreventive bioactivity of plant extracts from *Bidens pilosa*. Journal of Ethnopharmacology. 2004; 95(2-3): 409-19.
- Djeridane A, Yousfi M, Nadjemi B, Boutassouna D, Stocker P, Vidal N. Antioxidant activity of some Algerian medicinal plants extracts containing phenolic compounds. Food Chemistry. 2006; 97(4): 654-60.
- Eberlein M, Scheibner KA, Black KE, Collins SL, Chan-Li Y, Powell JD, Horton MR. Anti-oxidant inhibition of hyaluronan fragment-induced inflammatory gene expression. Journal of Inflammation. 2008; 5(1): 20.
- El-Masry S, Amer ME, Abdel-Kader MS, Zaatout HH. Prenylated flavonoids of *Erythrina lysistemon* grown in Egypt. Phytochemistry. 2002; 60(8): 783-7.
- El-Sayed MA, BaAbbad R, Balash A, Al-Hemdan NA, Softah A. The potential anti helicobacter pylori and antioxidant effects of *Artemisia Judaica*. Functional Foods in Health and Disease. 2013; 3(9): 332-40.
- 18. Erol-Dayi Ö, Pekmez M, Bona M, Aras-Perk A, Arda N. Total phenolic contents, antioxidant activities cytotoxicity of Three *Centaurea* Species: *C. calcitrapa* subsp. *calcitrapa*, *C. ptosimopappa C. spicata*. Free Radicals and Antioxidants. 2011; 1(2): 31-6.
- Essiett UA, Akpan EM. Proximate composition and phytochemical constituents of Aspilia africana (Pers) CD Adams and Tithonia diversifolia (Hemsl) A. Gray stems (Asteraceae). Bulletin of Environment, Pharmacology and Life Sciences. 2013; 2: 33-7.
- Freedman B, Nowak LJ, Kwolek WF, Berry EC, Guthrie WD. A bioassay for plant-derived pest control agents using the European corn borer. Journal of Economic Entomology. 1979; 72(4): 541-5.
- Gautam R, Saklani A, Jachak SM. Indian medicinal plants as a source of antimycobacterial agents. Journal of Ethnopharmacology. 2007; 110(2): 200-34.
- Halliwell B. Free radicals, antioxidants, and human disease: curiosity, cause, or consequence. The Lancet. 1994; 344(8924): 721-4.
- 23. Harborne, J. B., Phytochemical Methods, London. Chapman and Hall, Ltd. 1973: 49-188
- Iranbakhsh A, Ebadi M, Bayat M. The inhibitory effects of plant methanolic extract of *Datura stramonium* L. and leaf explant callus against bacteria and fungi. Global Veterinaria. 2010; 4(2): 149-55.
- Jamshidi-Kia F, Lorigooini Z, Amini-Khoei H. Medicinal plants: Past history and future perspective. Journal of Herbed Pharmacology. 2018; 7(1): 1-7.
- 26. Jimoh FO, Oladiji AT. Preliminary studies on *Piliostigma thonningii* seeds: proximate analysis, mineral composition and phytochemical screening. African Journal of Biotechnology. 2005; 4(12).
- 27. Joshi B, Lekhak S, Sharma A. Antibacterial property of different

- medicinal plants: *Ocimum sanctum, Cinnamomum zeylanicum, Xanthoxylum armatum* and *Origanum majorana*. Kathmandu University Journal of Science, Engineering and Technology. 2009; 5(1): 143-50.
- Kant MR, Jonckheere W, Knegt B, Lemos F, Liu J, Schimmel BC, Villarroel CA, Ataide LM, Dermauw W, Glas JJ, Egas M. Mechanisms and ecological consequences of plant defence induction and suppression in herbivore communities. Annals of Botany. 2015; 115(7): 1015-51.
- Khafagy SM, El-Din AS, Jakupovic J, Zdero C, Bohlmann F. Glaucolidelike sesquiterpene lactones from *Artemisia judaica*. Phytochemistry. 1988; 27(4): 1125-8.
- Khasawneh MA, Elwy HM, Fawzi NM, Hamza AA, Chevidenkandy AR, Hassan AH. Antioxidant activity, lipoxygenase inhibitory effect and polyphenolic compounds from *Calotropis procera* (Ait.) R. Research Journal of Phytochemistry. 2011; 5(2): 80-8.
- Kumar GS, Jayaveera KN, Kumar CK, Sanjay UP, Swamy BM, Kumar DV. Antimicrobial effects of Indian medicinal plants against acne-inducing bacteria. Tropical Journal of Pharmaceutical Research. 2007: 6(2): 717-23.
- Laurena AC, Den Truong V, Mendoza EM. Effects of condensed tannins on the in vitro protein digestibility of cowpea [Vigna unguiculata (L.) Walp.]. Journal of Agricultural and Food Chemistry. 1984; 32(5): 1045-8.
- 33. Leutner S, Eckert A, Müller WE. ROS generation, lipid peroxidation and antioxidant enzyme activities in the aging brain. Journal of Neural Transmission. 2001; 108(8-9): 955-67.
- Liu CZ, Murch SJ, El-Demerdash M, Saxena PK. Artemisia judaica L.: micropropagation and antioxidant activity. Journal of Biotechnology. 2004; 110(1): 63-71.
- 35. Mehraban, R., Lean Production. Tehran 2005, Jahane Farda pub.
- Miguel MG. Antioxidant and anti-inflammatory activities of essential oils: a short review. Molecules. 2010; 15(12): 9252-87.
- Mothana RA, Abdo SA, Hasson S, Althawab F, Alaghbari SA, Lindequist U. Antimicrobial, antioxidant and cytotoxic activities and phytochemical screening of some Yemeni medicinal plants. Evidence-Based Complementary and Alternative Medicine. 2010; 7(3): 323-30.
- 38. Obadoni BO, Ochuko PO. Phytochemical studies and comparative efficacy of the crude extracts of some haemostatic plants in Edo and Delta States of Nigeria. Global Journal of Pure and Applied Sciences. 2002; 8(2): 203-8.
- Okwu, D.E. and M.E. Okwu, Chemical composition of spondias mombin plants. Journal of Sustainable Agriculture and the Environment, 2004. 6: 140-147.
- Pandey AK, Singh P. The genus Artemisia: A 2012–2017 literature review on chemical composition, antimicrobial, insecticidal and antioxidant activities of essential oils. Medicines. 2017; 4(3): 68-83.
- Rashidi A, Mousavi B, Rahmani MR, Rezaee MA, Hosaini W, Motaharinia Y, Davari B, Zamini G. Evaluation of antifungal effect of Lavandula officinalis, Salvia officinalis L., Sumac, Glycyrrhiza glabra, and Althaea officinalis extracts on Aspergillus niger, Aspergillus fumigatus, and Aspergillus flavus species. Journal of Medicinal Plants Research. 2012; 6(2): 309-13.
- 42. Ravid U, Putievsky E, Katzir I, Carmeli D, Eshel A, Schenk HP. The essential oil of *Artemisia judaica* L. chemotypes. Flavour and Fragrance Journal. 1992; 7(2): 69-72.
- 43. Rios JL, Recio MC. Medicinal plants and antimicrobial activity. Journal of Ethnopharmacology. 2005; 100(1-2): 80-4.
- 44. Kordali S, Kotan R, Mavi A, Cakir A, Ala A, Yildirim A. Determination of the chemical composition and antioxidant activity of the essential oil of *Artemisia dracunculus* and of the antifungal and antibacterial activities of Turkish *Artemisia absinthium*, *A. dracunculus*, *Artemisia santonicum*, and *Artemisia spicigera* essential oils. Journal of Agricultural and Food Chemistry. 2005; 53(24): 9452-9458.
- Sadasivam, S. and A. Manickam, Biochemical Methods, 3rd Edition 2008., New Age International Publishers, New Delhi, India.
- Saleh NA, El-Negoumy SI, Abou-zaid MM. Flavonoids of Artemisia judaica, A. monosperma and A. herba-alba. Phytochemistry. 1987; 26(11): 3059-64.
- 47. Salem N, et al., Antioxidant activity of Silybum marianum and Ajuga iva natural dyes. International Journal of Control, Energy and Electrical Engineering, 2016; 3(2): 6-12.
- 48. Sharma YK, Davis KR. The effects of ozone on antioxidant responses in plants. Free Radical Biology and Medicine. 1997; 23(3): 480-8.
- 49. Sitapha O, Elisee KK, Joseph DA. Antifungal activities of Terminalia

- ivorensis A. Chev. bark extracts against *Candida albicans* and *Aspergillus fumigatus*. Journal of Intercultural Ethnopharmacology. 2013: 2(1): 49-52.
- Stray, F. The natural guide to medicinal herbs and plants 1998 (pp.12-16). Tiger Books International, London.
- 51. Su HC, Horvat R. Isolation, identification, and insecticidal properties of *Piper nigrum* amides. Journal of Agricultural and Food Chemistry. 1981; 29(1): 115-8.
- Alghanem SM, El-Amier YA. Phytochemical and Biological Evaluation of *Pergularia tomentosa* L. (Solanaceae) Naturally Growing in Arid Ecosystem. International Journal of Plant Science and Ecology. 2017; 3: 7-15.
- 53. Thorat B. Chemical extraction and biomedical importance of secondary organic metabolites from plants—A review. Journal of Biomedical and Therapeutic Sciences. 2018; 5(1): 9-42.
- Tiku AR. Antimicrobial Compounds and Their Role in Plant Defense. InMolecular Aspects of Plant-Pathogen Interaction 2018 (pp. 283-307). Springer, Singapore.
- 55. Van Buren JP, Robinson WB. Formation of complexes between protein and tannic acid. Journal of Agricultural and Food Chemistry. 1969; 17(4): 772-777.

- 56. Watson RR, Preedy VR, editors. Bioactive foods in promoting health: fruits and vegetables. Academic Press; 2009 Nov 24.
- El-Amier YA, Al-Hadithy ON, Abdulhadi HL, Fayed EM. Evaluation of antioxidant and antimicrobial activities of *Euphorbia terracina* L. from Deltaic Mediterranean coast, Egypt. Journal of Natural Products and Resources, 2016b; 2(2): 83-85.
- El-Amier YA, Abdullah TJ. Evaluation of nutritional value for four kinds of wild plants in Northern sector of Nile Delta, Egypt. Open Journal of Applied Sciences. 2015; 5(07): 393-402.
- 59. El-Amier YA, Abdelghany AM, Abed Zaid A. Green synthesis and antimicrobial activity of *Senecio glaucus*-Mediated silver nanoparticles. Research Journal of Pharmaceutical, Biological and Chemical Sciences. 2014; 5(5): 631-42.
- El-Amier YA, Al-hadithy ON, Abdullah TJ. Antioxidant and antimicrobial activity of different extracts obtained from aerial parts of *Urospermum* picroides (L.) F.W. from Egypt. Journal of Advanced Chemistry and Sciences, 2016a; 2(3): 299-301.
- 61. Zeng HY, Alan AR, Saxena PK. Evaluation of in vitro shoots of *Artemisia judaica* for allelopathic potential. Acta Physiologiae Plantarum. 2009; 31(6): 1237-48.

8 J Exp Sci • 2019 • Vol 10