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Radon Levels in different types of Plants with Medicinal Properties

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Abstract

In present study, natural levels of radon-222 in forty selected herbs with medicinal properties present in many stores throughout Iraq were measured to establish any potential radiation hazards. The Solid State Nuclear Track Detectors SSNTDs (CR-39) technique was used to determine these natural levels of radiation. The findings indicate that the radon concentrations ranged from 10.6602.07 Bq/m³ to 53.3034.64 Bq/m³ with an average 26.5373.21Bq/m³. These radon concentration values were lower than those reported in literature. These results show that consumption of the studied plants would impose no health threat to the consumers.

Keywords: Herbs; Medicinal plants; Radon concentration; Nuclear track detector; CR-39.

Introduction

Iraq has a long history and tradition of using herbs and other medicinal plants in traditional therapy which preceded the use of modern drugs and other antibiotics [1,2]. For thousands of years people have used and depended on herbs to treat many diseases. Presently, modern medicine and traditional or alternative medicine [3-5] are not exclusive, rather complementary to each another.

Radon-222 is a noble gas formed from radium (²²⁶Ra) and has a half-life of 3.8 days. Radon-222 can be emitted from the earth, rocks, as well as construction materials, and it can accumulate with its short-lived progeny in the atmosphere within the residences [6]. Exposure and inhalation of ²²²Rn for a short time period may lead to lung cancer. As a result of natural radioactive decay in the soil radon isotope particles, ²²²Rn are released from the soil particles escaping into the atmosphere. The radon release rate from the soil is known as the radon emanation rate or the radon exhalation rate. Radon exhalation is an intricate phenomenon depending on a number of parameters such as soil morphology, radium content in the soil, temperature, atmospheric pressure, soil moisture, rainfall and soil particle size [7]. The Solid State Nuclear Track Detectors or SSNTD's are considered to be most used devices for radon concentration measurements in the ecological fields. They are widely used in detecting and measuring radioactivity in geological samples and studying the influence of pollution in the dwellings [8]. The CR – 39 plastic track detectors were used for the evaluation of radon concentration in different types of herbs used in this study.

Materials and Methods

Sample Collection and Preparation

In September of 2015, forty different samples of medicinal plants were collected from the local markets from various places in Najaf City, Iraq. Samples were classified into groups as shown in table 1. The cursor in front of each sample represents the sample code, trade name, scientific name, part used and country of origin. The samples were prepared by drying them

for two to four days at the temperature from 42 to 44°C to eliminate absorbed moisture and obtain actual, dry weight. The dried samples were ground to a fine powder of equal size particles by using a blender. By using a highly sensitive scale with a tolerance $\pm 0.01\%$, we measured twenty grams of each individual sample for further analysis. Then the samples were placed in containers. Before use, containers were washed with dilute hydrochloric acid and rinsed with distilled water and assigned a code specific to each individual sample.

Table 1. List of herbs used in the study

No	Sample code	Trade name	Scientific name	Part used	Origin
1	H1	Senna	<i>Cassia senna L.</i>	Leaves	Saudi Arabia
2	H2	Safflower	<i>Carthamus tinctorius</i>	Flowers	Iran
3	H3	Ziziphus	<i>Ziziphus spina-Christi L.</i>	Leaves	Iraq
4	H4	Hops	<i>Humulus lupulus L.</i>	Peduncle	Iran
5	H5	Peppermint	<i>Mentha piperita L.</i>	Leaves	Iraq
6	H6	Balanite	<i>Balanites aegyptica(L.) Del.</i>	Fruits	Egypt
7	H7	Aelchenan	<i>Anabasis spp.</i>	Leaves	Iraq
8	H8	Green tea	<i>Camellia sinensis</i>	Leaves	China
9	H9	Fenugreek	<i>Trigonella foenum-graecum L.</i>	Seeds	India
10	H10	Sweet marjoram	<i>Origanum majorana</i>	Aerial parts	Middle east
11	H11	Ginger	<i>Zingiber officinale</i>	Roots	India
12	H12	Greater plantain	<i>Plantago major L.</i>	Peel fruits & seeds	India
13	H13	Hawthorn	<i>Crataegus spp.</i>	Leaves	USA
14	H14	Chokecherry	<i>Prunus virginiana L.</i>	Seeds	Azerbaijan
15	H15	Myrtle	<i>Myrtus communis L.</i>	Leaves	Iraq
16	H16	White cedar	<i>Thuja occidentalis</i>	Fruits	Syria
17	H17	Rosemary	<i>Rosmarinus officinalis L.</i>	Aerial parts	Mediterranean sea
18	H18	Chicory	<i>Cichorium intybus L.</i>	Roots, Stalk & leaves	Iraq
19	H19	Chamomile	<i>Matricaria chamomilla L.</i>	Flowers	Syria
20	H20	Sage	<i>Salvia officinalis</i>	Leaves	India
21	H21	Maidenhair fern	<i>Adiantum capillus-veneris L.</i>	Leaves and Stalk	USA
22	H22	Black mustard	<i>Brassica nigra(L.) W.D.J. Koch</i>	Seeds	China
23	H23	Cyperus	<i>Cyperus esculentus</i>	Seeds	Egypt
24	H24	Hollyhock	<i>Alcea rosea L.</i>	Flowers	India
25	H25	Ginkgo	<i>Ginkgo biloba.</i>	Seeds	Iran
26	H26	Bay leaves	<i>Laurus nobilis</i>	Leaves	Syria
27	H27	Corn Mint or Bo He	<i>Mentha haplocalyx</i>	Aerial parts	India
28	H28	Black cumin	<i>Nigella sativa L.</i>	Seeds	India
29	H29	Roselle	<i>Hibiscus sabdariffa L.</i>	Flowers	Iraq
30	H30	Horse tail	<i>Equisetium arvense L.</i>	Aerial parts	Egypt
31	H31	African Rue	<i>Ruta chalepensis L.</i>	Aerial parts	Saudi Arabia
32	H32	Flax	<i>Linum usitatissimum L.</i>	Seeds	Iran
33	H33	Garden Angelica/ Stout bien	<i>Angelica archangelica L.</i>	Each herb	China
34	H34	Yarrow	<i>Achillea millefolium</i>	Aerial parts	Iran
35	H35	Nutgrass	<i>Cyperus rotundus L.</i>	Roots and leaves	Saudi Arabia
36	H36	Colocynth	<i>Citrullus colocynthis (L) Shradc</i>	Fruits	Iraq
37	H37	Primrose	<i>Primula vulgaris L.</i>	Flowers	west Asia
38	H38	Borage	<i>Borago officinales</i>	Flowers	Iran
39	H39	Coltsfoot	<i>Tassilago farfara</i>	Leaves and flowers	North Asia
40	H40	Rose of Jericho	<i>Anastatica hierochuntica L.</i>	Branches	Palestine

Laboratory Procedure

Measurements were carried out upon 30 days after reaching the radiation equilibrium. Beacon covers were removed rapidly to prevent outside air from entering and changing the atmosphere in the cans. The nuclear detector CR-39 with dimensions of 1 cm² and 1 mm thick was placed at the middle of the underside of the cover and affixed with an adhesive tape. The edges of the cover were taped and sealed to prevent radon from leaking. The CR-39 detector recorded the presence and effects of alpha particles which resulted from the dissolution of radon gas. The distance between the surfaces of the sample and reagent was 5cm and the sample height was 2 cm as shown in figure 1. We applied the long-term method of 90 day exposure before removing the reagents exposing them to the chemical skimming procedure.

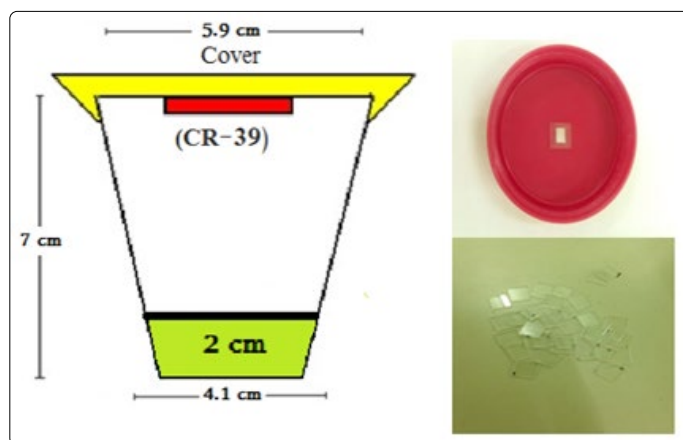


Figure 1. Schematic representation of the plastic container showing the position of the CR-39 detector and the sample tested.

Chemical Etching and Microscopic Scanning

Detectors were removed and etched in a 6.25N aqueous solution of NaOH. The detectors were placed inside Pyrex and linked with a wire. The Pyrex was placed inside a water bath. Following the standard protocol of keeping them in the bath at 70°C for 7 hours, detectors were rinsed with distilled water and allowed to air-dry than placed in a plastic box [9].

The tracks recording the effect of alpha particles at the surface of the CR-39 nuclear detectors were observed by using novel optical microscope at 400x magnification as shown in figure 2. A total of five optic view fields were selected for taking the readings for each individual sample.

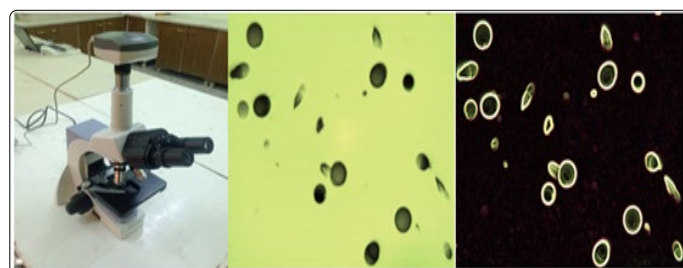


Figure 2. The effects of alpha particles chemical etching as seen at 400x by optical microscope.

Calculation of Radon concentrations

The density of the tracks ρ in the detectors was calculated according to the following equation [10].

$$\rho = \frac{N_{average}}{A} \dots \dots \dots (1)$$

Where, ρ is track density (Track /cm²), N is an average of total tracks (Track) and A is an area of a view field (cm²).

Radon concentration (C_{Rn}) in Bq/m³ unit are calculated by the following equations [11,12].

$$C_{Rn} = \frac{\rho}{k T} \dots \dots \dots (2)$$

Where, k : is the calibration factor in terms of (track.cm⁻²/Bq.d.m⁻³) which is the same value as reported in many works [11-14]. T is the exposure time (d).

The value for radon activity (A_{Rn}) and specific radon activity ($S.A_{Rn}$) can be found based on radon concentrations, volume of container (V) and mass of a sample (m) as it follows: [8,15-17]

$$A_{Rn} = C_{Rn} V \dots \dots \dots (3)$$

$$S.A_{Rn} = \frac{A_{Rn}}{m} \dots \dots \dots (4)$$

Results and Discussion

Table 2 presents radon concentration and specific activity for medicinal plant samples originated in different countries but obtained at the local markets. Our study shows that the lowest value of radon concentration was found in H6 Egypt sample, which was (10.6602.07 Bq/m³), while the highest average value was found in H35 Saudi Arabia sample, having values of (53.3034.64 Bq/m³) as shown in figure 3. The same sample from Saudi Arabia had shown specific radon activity. The specific radon activity varied from 0.003 ± 0.0003 Bq/kg in H₂O samples to 0.7790.085 Bq/kg in H30 samples with an average 0.2830.035 Bq/kg. These results indicate that the reasons for variation of radon concentration in different medicinal plant samples are the result of chemical reaction by which radon transfer occurs from the soil solids into the water solution within the soil becoming absorbed by the roots and translocated throughout the plants. The uptake of these radio nuclides from the soil solution is controlled by plant physiology. Our findings are similar and very close to the reported results cited in the literature as presented in table 3.

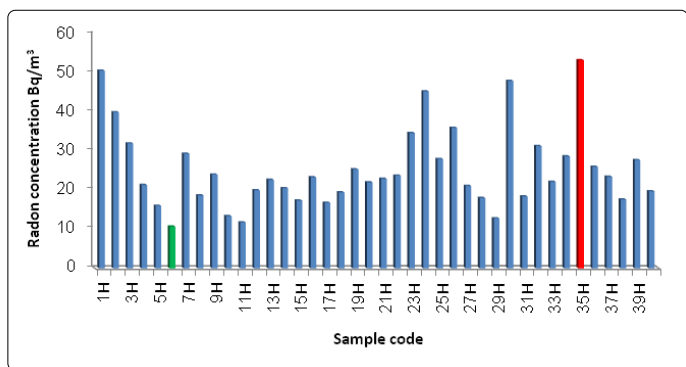


Figure 3. Radon concentration in medicinal plants.

Table 2: The radon concentration and specific radon activity in medicinal plants.

Sample code	C_{Rn} (Bq/m ³)	$S.A_{Rn}$ (Bq/kg)
H1	50.6384.52	0.4700.042
H2	39.9774.01	0.6490.085
H3	31.9823.59	0.0310.277
H4	21.3212.93	0.0290.213
H5	15.9912.54	0.0300.188
H6	10.6602.07	0.0080.046
H7	29.3163.44	0.0230.200
H8	18.6562.74	0.0230.161
H9	23.9863.11	0.0220.173
H10	13.3252.32	0.0270.157
H11	11.7262.17	0.0170.095
H12	19.9882.84	0.0280.199
H13	22.6533.02	0.0490.368
H14	20.5212.87	0.0240.177
H15	17.3232.64	0.0260.173
H16	23.3203.06	0.0210.159
H17	16.7902.60	0.0330.218
H18	19.4552.80	0.0600.421
H19	25.3193.19	0.0590.470
H20	21.9872.98	0.00030.003
H21	22.9203.04	0.0490.372
H22	23.7203.09	0.0260.205
H23	34.6473.74	0.0280.264
H24	45.3074.27	0.0460.490
H25	27.9843.36	0.0310.259
H26	35.9793.81	0.0300.292
H27	21.0542.91	0.0340.248
H28	17.9892.69	0.0200.137
H29	12.7922.27	0.0130.075
H30	47.9734.40	0.0710.779
H31	18.3892.72	0.0350.239
H32	31.3153.55	0.0270.239
H33	22.1202.98	0.0350.261
H34	28.6503.40	0.0550.465
H35	53.3034.64	0.0240.277
H36	25.9853.24	0.0280.225
H37	23.4533.07	0.1000.762
H38	17.5902.66	0.0430.285
H39	27.7173.34	0.0620.514
H40	19.7222.82	0.0200.142
Max	53.3034.64	0.7790.085
Min	10.6602.07	0.00030.003
Average	26.5373.21	0.2830.035

Table 3. Our results showing radon concentration compared to findings cited in literature.

Country	Average of radon concentrations (Bq/m ³)	Reference
Iraq(Bagdad)	19.2	[8]
Morocco	10.2	[17]
Iraq(Najaf)	26.5	This work

Conclusion

The forty herb samples collected from the local markets from various places in Iraq were evaluated for any potential radiation hazards by using SSNTDs (CR-39 detector). Our research indicates that the highest concentration of radon is found in roots and leaves, while the lowest radon concentration value is found in fruits. The radiological study shows that consumption of the studied medicinal plants would impose no health threat to the consumers.

References

1. Rukangira E. The African Herbal Industry: Constraints and Challenges. 2012.
2. Kabir OA, Olukayode O, Chidi EO, Christopher CI, Kehinde AF. Screening of crude extracts of six medicinal plants used in South-West Nigerian unorthodox medicine for anti-methicillin resistant *Staphylococcus aureus* activity. *BMC Complement Altern Med*. 2005; 5(1). doi: 10.1186/1472-6882-5-6
3. Priya KS, Gnanamani A, Radhakrishnan N, Babu M. Healing potential of *Datura alba* on burn wounds in albino rats. *J Ethnopharmacol*. 2002; 83(3): 193–199.
4. Steenkamp V, Mathivha E, Gouws MC, Rensburg CEJ. Studies on antibacterial, antioxidant and fibroblast growth stimulation of wound healing remedies from South Africa. *J Ethnopharmacol*. 2004; 95(2-3): 353-357. doi: 10.1016/j.jep.2004.08.020
5. Patel M, Coogan MM. Antifungal activity of the plant *Dodonaea viscosa* var. *angustifolia* on *Candida albicans* from HIV-infected patients. *J Ethnopharmacol*. 2008; 118(1): 173-176. doi: 10.1016/j.jep.2008.03.009
6. Rawat A, Jojo PJ, Khan AJ, Tyagi RK, Rajendra P. Radon exhalation rate in building materials. *International Journal of Radiation Applications and Instrumentation. Part D. Nuclear Tracks and Radiation Measurements*. 1991; 19(1-4): 391-394. doi: 10.1016/1359-0189(91)90223-5
7. Khan MS, Srivastava DS, Azam A. Study of radium content and radon exhalation rates in soil samples of northern India. *Environmental Earth Sciences*. 2012; 67(5): 1363-1371. doi: 10.1007/s12665-012-1581-7
8. Dawser HG. Measurement of Radon Concentration in Henna Plant Using Etched Track Detectors. *Al-Mustansiriyah J Sci*. 2011; 22(1): 139-144.
9. Mohammad AM, Ahamed IK, Ahmed YS. Measurement of Uranium Concentration in Some Soil Samples in Jalawla'a City Using CR-39 Detector. *Journal of Al-Nahrain university*. 2013; 16(1): 112-116.
10. Ali AA, Hamidawi A, Afnan AH. Radiation Hazards Due to Radon Concentrations in Dwellings of Kufa Technical Institute, 'Iraq. *Physics International*. 2016; 7(1): 28-34. doi: 10.3844/pisp.2016.28.34
11. Al-Kofahi MM, Khader BR, Lehlooh AD, Kullab MK, Abumurad KM, Al-Bataina BA. Measurement of radon 222 in Jordanian dwellings. *International Journal of Radiation Applications and Instrumentation. Part D. Nuclear Tracks and Radiation Measurements*. 1992; 20(2): 377-382.
12. Rasas M. Measurement of Radon and Its Daughter's Concentrations In Indoor and Outdoor Throughout Gaza Strip. *Journal of the Association of Arab Universities for Basic and Applied Sciences*. 2003; 11(1): 21-26. doi: 10.1016/j.jaubas.2011.10.003
13. Mayya YS, Eappen KP, Nambi KSV. <https://academic.oup.com/rpd/article-abstract/77/3/177/1649504?redirectedFrom=fulltext> *Radiation Protection Dosimetry*. 1998; 77(3): 177–184.
14. Sersawi M. Study of the chronic radiation exposure situation in Gaza. Islamic University of Gaza in Physics. 2007.
15. Yousef HA, Saleh GM, El-Farrash AH, Hamza A. Radon exhalation rate for phosphate rocks samples using alpha track detectors. *Journal of Radiation Research and Applied Sciences*. 2016; 9(1): 41-46. doi: 10.1016/j.jrras.2015.09.002
16. Tykva R, Jozef S. Low-level environmental radioactivity: sources and evaluation. CRC Press, USA. 1995.
17. Oufni L, Manaut N, Taj S, Manaut B. Determination of Radon and Thoron Concentrations in Different Parts of Some Plants Used in Traditional Medicine Using Nuclear Track Detectors. *American Journal of Environmental Protection*. 2013; 1(2): 34- 40.