



# Green synthesis of *Abutilon indicum* (L) derived iron oxide (FeO) nanoparticles with excellent biological, anticancer and photocatalytic activities

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## ABSTRACT

Cancer is one of the most common causes of death all over the world. Although there are many treatments available none of them are without side-effect and most of them are very expensive. Similarly, Inflammation is one of another common medical condition worldwide but most of the therapies available are not environment and pocket friendly. Thus, the aim of our study was to synthesize environmentally friendly, cost-effective FeO NPs with significant anticancer, anti-inflammatory, antibacterial properties. The green synthesis of Iron Oxide Nanoparticles (FeO NPs) using plant extract is one of the environmentally friendly, cost-effective, and a secure alternative to physical or chemical synthesis methods. In the present study we used *Abutilon indicum* leaf extract to synthesize the Ai-FeO NPs and performed various experiments to confirm the therapeutic potential of green synthesized Ai-FeO nanoparticles. In this method, an aqueous plant leaf extract (*A. indicum*: Ai) was used as a reducing and stabilizing agent for the synthesis of FeO nanoparticles. The synthesized Ai-FeO NPs were characterized by the ultraviolet-visible (UV-Vis) absorbance of the surface at a resonance band 415 nm. Fourier transform infrared (FTIR) spectroscopy study confirmed the inherent bioactive functional groups as capping and stabilizing agents for Ai-FeO NPs. The X-ray powder diffraction (XRD) spectra revealed crystalline nature of the NPs. Further, the synthesized NPs have been isolated in highly stable, crystalline nature with spherical shape having size 10–95 nm. In addition, the green synthesized Ai-FeO-NPs demonstrated dose-dependent antimicrobial, antioxidant, anti-inflammatory, and anticancer properties. The findings of this study suggested that *A. indicum* leaf extract mediated FeO nanoparticles could be potentially used as a therapeutic agent for many human diseases and breast cancer.

## 1. Introduction

According to World Health Organization, cancer is one of the leading causes of death, and the death rate is approximately 9 million in 2018 [1]. However, middle- and low-income nations account for 70 % of cancer deaths worldwide [1,2]. Inflammation, another medical

condition, is among some of the most common medical issues worldwide. Fortunately, inflammation is an essential part of the healing process and maintains cellular function. However, acute, and chronic inflammations that are recognized could be problematic if not treated early [3]. The acute inflammation symptoms include redness, swelling, and pain inside tissues and joints [4]. Since both of the above-mentioned

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diseases are very common across the globe, there is an urgent need for new treatments methods that involve minimum cost and low or no side effects.

In recent years, nanotechnology has emerged as an important area of research in the fields of medicine and health. In the past few years, nanomedicine has revolutionized health care in many ways, and has many opportunities in a wide variety of sectors and scientific fields [5]. Nowadays, physical, chemical, and green methods are used for metallic nanoparticles synthesis. The use of chemical and physical methods comprises the formation of toxic chemical, high pressure, radiation, and involves costly equipment, results in a large quantity of chemical waste as a by-product. These waste chemical by-products could be hazardous to the environment [6,7]. Green synthesis of nanoparticles by using plant parts through biological methods is one of the very effective techniques, which results in higher yield than the other related techniques [8]. Plant components in the green synthesis play a crucial role as reducing and stabilizing agents along with other biochemical properties [9–11]. The green synthesis of metal and metal oxide nanoparticles is a novel method that avoids harmful chemicals, high cost and environmental contamination [12]. As a result, researchers are focussing on green approaches for producing nanoparticles from plants, microbes, and other biomaterials [8–13].

Plant extracts are easier way to synthesize metal oxide nanoparticles than microbial cultures because plant extracts don't require sterilized conditions to grow [10,11]. Usually, plant extracts contain phytochemicals including sugars, and several secondary metabolites such as polyphenols, flavonoids, terpenoids, phenolic acids, alkaloids, and protein, which causes the reduction and stabilization of metal nanoparticles [10]. Green synthesized NPs are promising antibacterial and anticancer agents and are safer, cost-effective medications than other commercial drugs [2,14]. Recent efforts have been focused on synthesizing environmentally friendly NPs such as Ag, Au, Cu, CuO, ZnO, Se and Fe, which have been integrated into a variety of biological activities [15,16]. Among the various metal nanoparticles, scientists have been paying significant attention to FeO-NPs due to its most appealing properties as a multifunctional material. FeO-NPs have received extensive research attention in a variety of fields, including plant growth regulators, sensors, photocatalysts, fine ceramics, water treatment, pigments, electrochemical cells, and anticorrosive chemicals [17–21]. The FeO NPs have a wide variety of commercial applications including, biomedicine, vaccines, diagnostics, radiology, pathogens, enzymes, and cosmetics [22]. Further, these FeO NPs are typically utilised for many purposes, such as the removal of heavy metals, dyes, and antibiotics from water sources, as well as in the biomedical area for targeted drug delivery to the cancer cells [23,24]. Due to their propensity to generate highly reactive oxygen species, iron oxide nanoparticles have been effective against a wide range of pathogenic bacterial strains and fungi [16]. Recently, researchers are focussing on green synthesis FeO NPs from various plant parts, for example the leaf extract of *Rhamnus triquetra* [25], *Platanus orientalis* leaf extract [26], *Madhuca indica* leaf extract [27], fruit extract of *Cynometra ramiflora* [28], *Carica papaya* leaf extract [20], *Hibiscus rosa sinensis* flower extract [14] and *Punica graatum* seeds extract [29], etc.

In the present study, FeO NPs were synthesized by green chemistry approach from *Abutilon indicum* (Malvaceae) plant parts which contains a variety of phytochemical such as alkaloids, proteins and amino acids, flavonoids, steroids, glycosides, saponins. The crude leaf extract of this plant had significant antioxidant, antibacterial, and anticancer activity [30–32]. Although *A. indicum* leaf extract is already reported to be used as reducing agent in the previous research to synthesize silver NPs [33], ZnO NPs [34], CuO NPs [35], MnO NPs [36], and Ni nanoparticles [37], but there are only few reports on the synthesis of FeO NPs from *A. indicum* leaf extract and their biological and photocatalytic properties.

Therefore, the present study focused on green synthesis of FeO NPs by using *A. indicum* leaf extract. Physical and chemical properties of the

synthesized FeO NPs were analyzed using UV–visible (UV–Vis) spectroscopy, XRD, FTIR, HR-TEM, EDAX, Zeta potential analysis. Also, we evaluate their antioxidant, antibacterial, anti-diabetic, anti-inflammation, and anticancer activity. Hence, this research not only demonstrates the green synthesis of FeO NPs but also illustrates the multifunctional application of such synthesized FeO NPs.

## 2. Materials and methods

### 2.1. Chemicals

Ferric acetate basic (hydrous), nutrient agar and broth, 2,2-diphenyl-1-picrylhydrazyl (DPPH), hydrogen peroxide, 2-Azino-bis (3-ethylbenzo-thiazoline-6-sulphonic acid) diammonium salt (ABTS), ascorbic acid, Acridine Orange/Ethidium bromide staining kit (AO/EB), 3-(4,5-dimethylthiazolyl-2-yl)-2, sodium nitroprusside, 5-diphenyl tetrazolium bromide (MTT).

### 2.2. Plant extracts preparation

*Abutilon indicum* leaf extracts were prepared according to the methods demonstrated by Venkatesan et al. [38] with slight modifications. 150 mL of deionized water and 5 g of *Abutilon indicum* leaf powder were combined and heated to 70 °C for one hour. Then the extracts were cooled at room temperature and separated with Whatman No. 1 filter paper. At the end the extracted solution was used to produce nanoparticles.

### 2.3. Green chemistry synthesis and characterization of Ai-FeO NPs

For the synthesis of Ai-FeO NPs, Abbasi et al. [23] method was adopted with slight modification. 100 mL of *A. indicum* leaf extract and 6 g of ferric acetate basic (hydrous) were combined to produce FeO NPs. The mixture was kept on a hot plate and heated continuously for two hours at 70 °C. The color of the solution changed from yellow to blackish after 2 hr, indicating the formation of FeO NPs. It was then cooled at room temperature. The powder at the bottom was retained, once the solution was centrifuged for 10 min at 12,000 rpm, Further, it was washed thrice with distilled water to remove all unreacted biological species from the residue. In the next step, the samples were washed with ethanol and calcinated for three hours at 400 °C to obtain the Ai-FeO NPs.

The green synthesized iron oxide nanoparticles were analyzed by UV–VIS light spectra (Jasco UV-650, Japan). The synthesized materials' purity and crystalline phase were examined using a Model D8, BRUKER AXS, and Cu K radiation ( $\lambda = 0.15425$  nm), in the range of 20° to 80°. The sizes and structure of Ai-FeO NPs were confirmed by transmission electron microscopy (TEM, Zeiss-EM10C). Additionally, their elemental mapping was examined using EDS (FE-SEM with EDS, Carl Zeiss- Sigma model, Germany), and stability using zeta potential with the help of Malvern analyser (Model- Nano-ZS90). FTIR (in the range The 400–4000  $\text{cm}^{-1}$ ) was used to characterize the capping agents of the leaf extract, while synthesizing Ai-FeO NPs.

### 2.4. Antioxidant activity of green synthesized FeO NPs

In vitro antioxidant assays including DPPH (2,2-diphenyl-1-picrylhydrazyl), reducing potential and Nitric oxide scavenging property of FeO NPs, Ascorbic acid at different dosage such as 20, 40, 60, 80, 100 and 120  $\mu\text{g/ml}$ , was determined by UV–VIS spectrophotometer using different wavelength such as 515 nm for DPPH, 700 nm for ABTS and 546 nm for NO assays [39,40].

### 2.5. Antibacterial activity of FeO NPs

The bactericidal potential of *A. indicum* leaf extract and Ai-FeO NPs

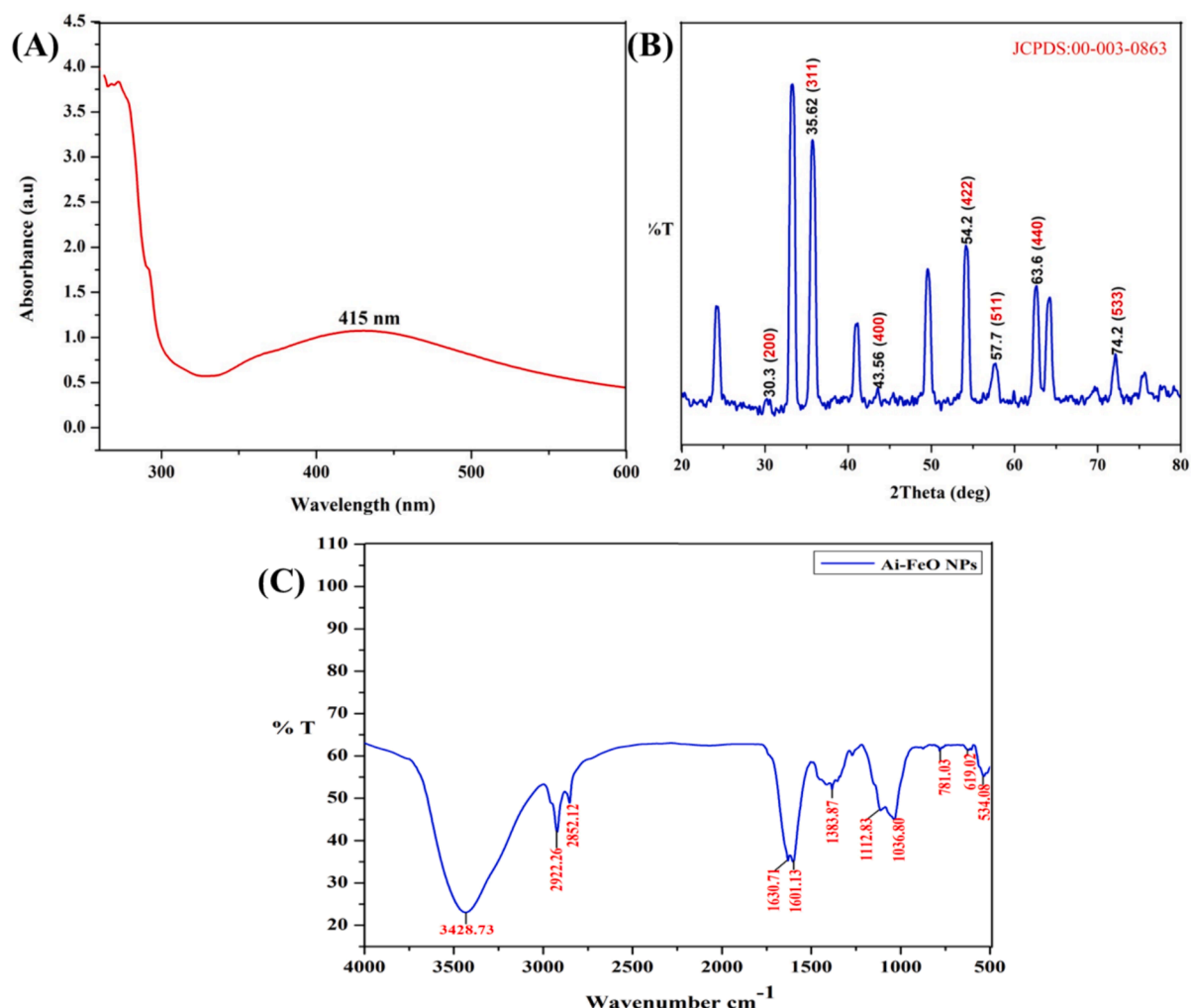


Fig. 1. (A) UV-vis spectrum of *Abutilon indicum*-synthesized FeO NPs; (B) XRD pattern of Ai-FeO NPs; (C) Fourier transform infrared (FTIR) spectroscopy of FeO NPs using *A. indicum*.

was assessed by using the previously developed agar well diffusion method [40,41]. Antibacterial activity was assessed using clinically isolated gram + ve bacteria (*S. aureus*, *B. subtilis*), as well as gram-ve (*E. coli*, *P. aeruginosa*) bacteria. 100  $\mu$ l of mature cultures that were 24 h old were swabbed onto the nutrient agar medium using an L-shaped rod. Further, the wells were created (6 mm) by sterile cork borer. The next step involved the study of various Ai-FeO NPs concentration (25, 50, 75, 100  $\mu$ g/ml) on different bacterial strains. After swabbing, the plates were incubated at 37  $^{\circ}$ C for 24 h, and their Zone of Inhibition (mm) values were periodically assessed.

## 2.6. In vitro anti-inflammatory activity

In vitro anti-inflammation assays were carried out by following the method of Rajakumar et al. [42] with slight modification. Herein, we examined 5 mL of 1 % bovine serum albumin and Ai-FeO NPs, and diclofenac at different dosage like 20, 40, 60, 80, 100 and 120  $\mu$ g/ml, and the activity was determined by UV-vis spectrophotometer using different wavelength such as 540 nm for  $\alpha$ -amylase, 660 nm for protein inhibitory assays.

## 2.7. Cytotoxic activity using MTT assay

MTT assay was used to demonstrate the cell viability of green synthesized Ai-FeO NPs, DOX. Breast cancer cells (MDA-MB-231) were

injected into the 96-well plates. Then the cancer cells treated with varying concentration of DOX, Ai-FeO NPs (20 to 120  $\mu$ g/ml) for 24 h at 37  $^{\circ}$ C. Following incubation, the OD was analyzed at 570 nm, and the IC<sub>50</sub> values were calculated [43].

### 2.7.1. Dual staining (AO/EB) and DAPI analysis of apoptosis

The apoptosis related morphological characteristic features were analyzed by staining the cells with acridine orange and ethidium bromide (AO/EB), as well as 4',6-Diamidino-2-phenylindole dihydrochloride (DAPI). We determined the IC<sub>50</sub> of Ai-FeO NPs after the cells were sown in six-well plates for 48 h. Then twenty micro liters of fluorescence dye were added after the well plates have been treated. Further, the cells were examined with fluorescent dyes under a fluorescence microscope [44].

### 2.8. Photocatalytic activities of methylene blue dye degradation

Photocatalytic activities of green synthesized Ai-FeO NPs were analyzed using methylene blue degradation in presence of sunlight radiations at different time intervals. As an inventory solution, methylene blue (1 mg) dye was dissolved in double distilled water (100 mL). 20 mg of green synthesized Ai-FeO NPs were distributed in 50 mL of methylene blue dye solution. A control was used without the inclusion of Ai-FeO NPs. Before exposing to irradiation, the reaction suspension was carefully mixed by using magnetic agitator for 30 min to confirm that the

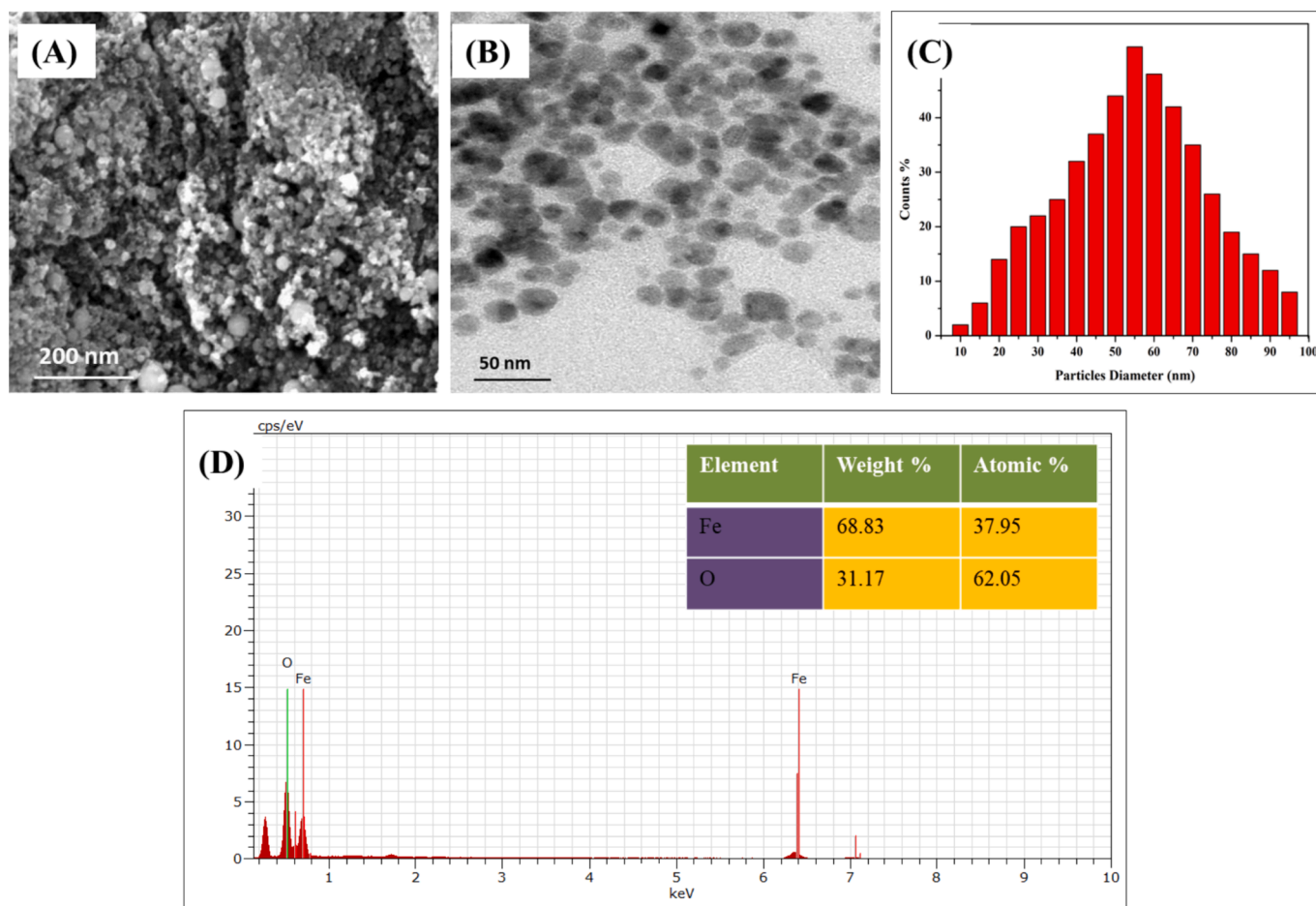


Fig. 2. (A) SEM image of *A.indicum*-synthesized FeO NPs (B) HR-TEM image of *A.indicum*-synthesized FeO NPs (C) Particle size distribution pattern of Ai-FeO NPs and (D) EDX analysis of *A.indicum*-synthesized FeO NPs.

working solution is homogeneous. The dispersion was then exposed to sunlight and monitored for different time intervals. Aliquots of 5 mL solutions were obtained and filtered at specific time intervals to examine the photocatalytic degradation of dye via UV spectra in the range 200–700 nm.

### 3. Statistical analysis

The ANOVA Tukey's HSD test was carried out using the IBM SPSS statistics software. Means were compared by the least significant difference (LSD) method at  $p < 0.05$ .

## 4. Results and discussion

### 4.1. 1. Green synthesis and characterization of Ai-FeO NPs

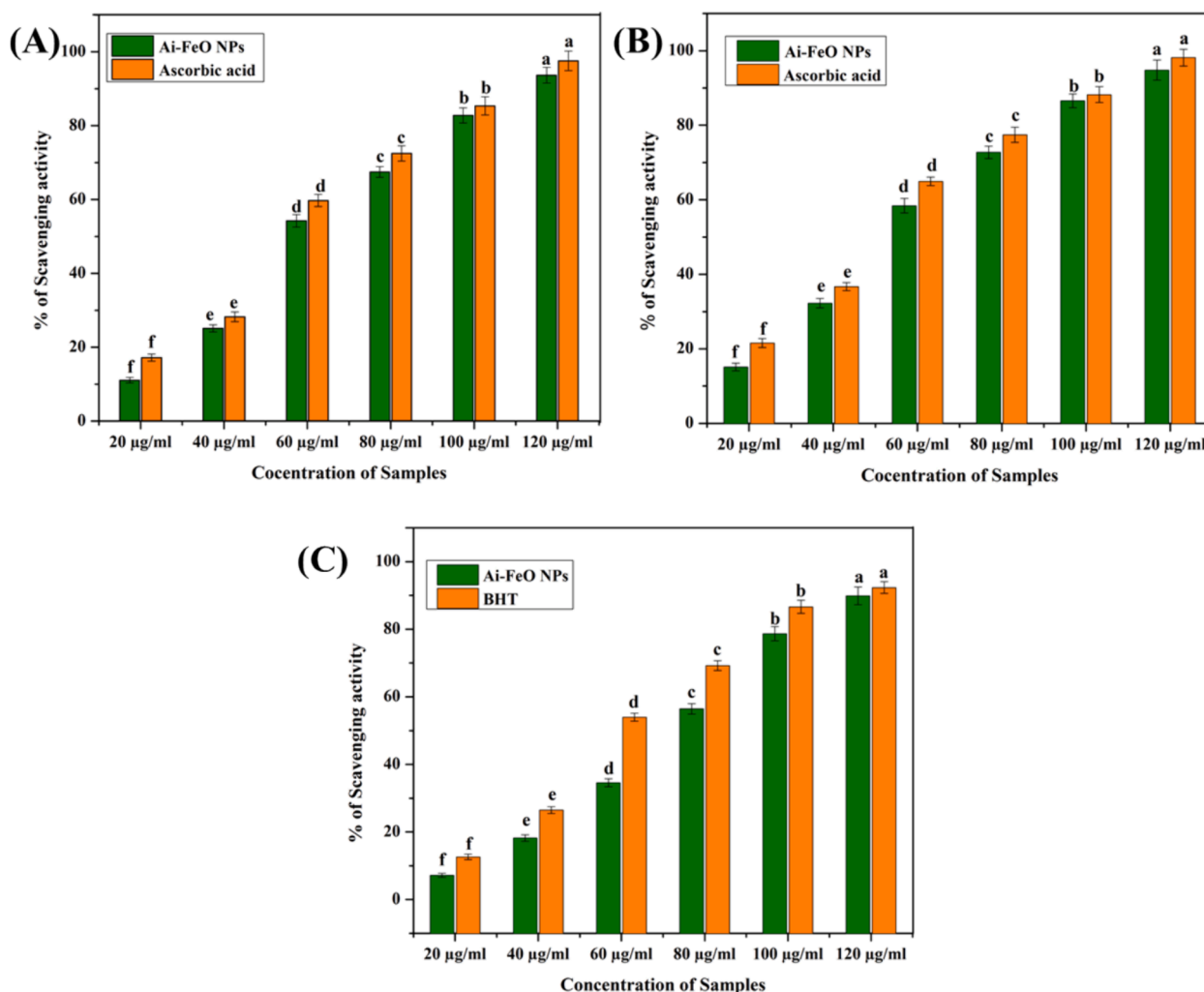
An environmentally friendly approach was used to synthesis FeO nanoparticles from *A. indicum* leaf extract. The green synthesized FeO NPs was preliminary confirmed by a distinctive transformation of the plant extract solution's color from yellow to a dark brown solution when the iron salt solution was introduced. *A. indicum* leaf extract contains active phytochemicals that reduce Fe ions to FeO NPs by the excitation of surface Plasmon resonances (SPR) on synthesized Ai-FeO NPs, resulting in Fe metal reduction [10,19]. Based on the spectra Fig. 1A, the peak surface plasmon resonance (SPR) bands at around 415 nm, which is in agreement with the previously reported SPR for FeO NPs using pomegranate seeds extract [18]. A similar study was conducted by Viju Kumar et al. (2018) [45] who measured the absorbance spectra of FeO

NPs in the 330–400 nm range.

The crystalline nature of the synthesized Ai-FeO NPs was determined by XRD analysis. The peak positions with  $2\theta$  values corresponding to  $30.3^\circ$ ,  $35.62^\circ$ ,  $43.56^\circ$ ,  $54.2^\circ$ ,  $57.7^\circ$ ,  $63.6^\circ$ , and  $74.2^\circ$  are associated with the indexed planes (200), (311), (400), (422), (511), (440), and (533), respectively (Fig. 1B). Undoubtedly, the strong and distinct peaks indicates that the  $\text{Fe}_2\text{O}_3$  nanoparticles produced by the reduction process using *A. indicum* leaf extract were of crystalline nature. According to the database of the Joint Committee on Powder Diffraction Standards (JCPDS No. 00–003–0863), these peaks indicate that the synthesized Ai-FeO NPs are of crystalline structures in nature. The results are nearly identical to those reported by other research associated with iron oxide nanoparticles [31,46].

The biomolecules present in *A. indicum* plant extract induce the reduction of Fe to Ai-FeO NPs which was confirmed using FTIR measurements. Fig. 1 (C) of FTIR spectra shows peaks at  $3428$ ,  $2922$ ,  $2852$ ,  $1630$ ,  $1601$ ,  $1383$ ,  $1112$ ,  $1036$ ,  $781$ ,  $619$  and  $534 \text{ cm}^{-1}$ . The broad peak at  $3428 \text{ cm}^{-1}$  was attributed to the primary amine and amide (O–H stretching). The peaks at  $2922$  and  $2852 \text{ cm}^{-1}$ , corresponds to C–H stretching in alkanes and carboxylic acid O–H stretching. The presence of secondary amine, amide is indicated by the medium spectral peaks at  $1630$ ,  $1601 \text{ cm}^{-1}$ . It is worth noting that the Fe–O bonds of magnetite have a characteristic vibration between  $619$  and  $500 \text{ cm}^{-1}$ . Our findings are also in agreement with earlier reports that show the presence of FeO NPs in peak ranging from  $400$  to  $624 \text{ cm}^{-1}$  [47].

The shape and size of the green synthesized Ai-FeO NPs by *A. indicum* leaf extract was shown in Fig. 2A, B. TEM image shows an excellent distribution of spherical Ai-FeO NPs with no aggregation. Furthermore,



**Fig. 3.** Percentage of DPPH scavenging activity of Ascorbic acid and Ai-FeO NPs; (B) ABTS Radical scavenging activity of Ascorbic acid and Ai-FeO NPs; (C) Nitric oxide scavenging activity of BHT and Ai-FeO NPs. The data represents the mean values of three independent experiments and are presented as mean  $\pm$  SD of the absorbance. Different letters indicated the significance differences (ANOVA followed by Tukey's HSD,  $\alpha = 0.005$ ).

**Table 1**

IC<sub>50</sub> scavenging activity of green synthesized Ai-FeO NPs, Ascorbic acid.

Assays	IC <sub>50</sub> value	
	Stranded	Ai-FeO NPs
DPPH	52.21 $\pm$ 1.22 <sup>a</sup>	60.41 $\pm$ 1.68 <sup>a</sup>
ABTS	47.27 $\pm$ 1.55 <sup>b</sup>	56.62 $\pm$ 2.04 <sup>b</sup>
NO	58.39 $\pm$ 1.46 <sup>c</sup>	75.14 $\pm$ 2.63 <sup>c</sup>

Values mean  $\pm$  SD indicates the replicates of three experiments.

\*Mean values within the column followed by the same ANOVA letter in superscript are significantly different at  $P < 0.05$  level.

imaging examination revealed that the size of the synthesized Ai-FeO NPs ranged from 10 to 95 nm as indicated in Fig. 2C. Recently, *Hibiscus rosa sinensis* flower extract was used to create spherical FeO NPs with a particle size 55 nm [14]. EDX analysis was used to analyze the quantitative elemental mapping of green produced Ai-FeO NPs (Seen in Fig. 2D). The data analysis revealed that the synthesized Ai-FeO NPs contains Fe, O, with weight percentages of 68.83, and 31.17 %, respectively. EDX results reveal that FeO NPs was successfully synthesized from *A. indicum*. According to EDX analysis, Fe and O made up the majority of the elements in the nanostructure.

#### 4.2. Radical scavenging activity of Ai-FeO NPs

The DPPH scavenging activity of the two samples (Ai-FeO NPs, Ascorbic acid) was conducted in triplicate. The DPPH free radical is considered to be a stable free radical that plays a critical role in the reduction of hydrogen or electrons from donors [48]. The Ai-FeO NPs ability to reduce DPPH was evaluated based on color change. The green synthesized Ai-FeO NPs demonstrated good scavenging performance but slightly low activity in comparison to the L-ascorbic acid. The percentage inhibition for different concentration (like 20, 40, 60, 80, 100 and 120 µg/ml) of Ai-FeO NPs and ascorbic acid samples was tested. Our results show a scavenging activity of 11.08  $\pm$  0.75, 25.13  $\pm$  0.98, 54.27  $\pm$  1.68, 67.46  $\pm$  1.44, 82.77  $\pm$  2.06 and 93.61  $\pm$  2.14 for Ai-FeO NPs. However, ascorbic acid was found to be 17.19  $\pm$  0.94, 28.26  $\pm$  1.32, 59.73  $\pm$  1.65, 72.48  $\pm$  2.07, 85.36  $\pm$  2.47 and 97.55  $\pm$  2.63 respectively which are seen in Fig. 3 (A). On the other hand, the IC<sub>50</sub> values for DPPH scavenging activity for Ai-FeO NPs and L-ascorbic acid were determined from the plotted regression graph to be 60 µg/ml and 52 µg/ml, respectively, (Table 1). Comparably, Younas et al. [49] highlighted that *Ixora finlaysoniana* extract derived Fe-Cu bimetallic nanoparticles exhibited DPPH radical scavenging activity at concentrations of 200–600 ppm respectively.

As a result of their reducing power assay, they can donate an electron and can inhibit the oxidation of intermediates in the lipid peroxidation

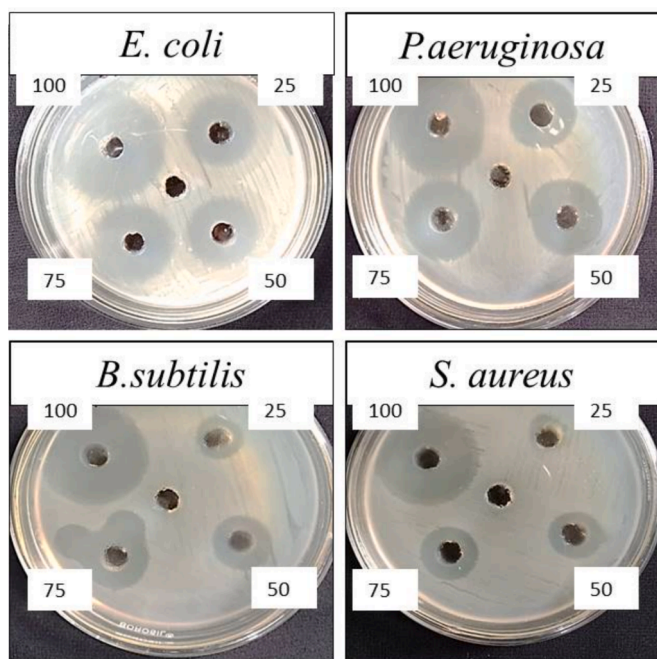


Fig. 4. Antibacterial activity *A.indicum* synthesized FeO NPs on *S. aureus*, *B. subtilis*, *E.coli*, and *P. aeruginosa*.

Table 2

Antimicrobial activity of green synthesized Ai-FeO NPs and *A. indicum* leaf extract.

Samples	Bacteria	Zone of inhibition (mm)			
		Concentration of samples			
		25 µg/ml	50 µg/ml	75 µg/ml	100 µg/ml
Ai-FeO NPs	<i>E. coli</i>	8.12 ± 0.72 <sup>d</sup>	11.31 ± 0.93 <sup>c</sup>	16.68 ± 0.98 <sup>b</sup>	19.25 ± 1.08 <sup>a</sup>
	<i>P. aeruginosa</i>	5.48 ± 0.67 <sup>d</sup>	8.62 ± 0.47 <sup>c</sup>	11.16 ± 0.85 <sup>b</sup>	16.56 ± 0.64 <sup>a</sup>
	<i>S. aureus</i>	3.51 ± 0.72 <sup>d</sup>	6.92 ± 0.21 <sup>c</sup>	8.21 ± 0.66 <sup>b</sup>	12.63 ± 1.07 <sup>a</sup>
	<i>B.subtilis</i>	4.68 ± 0.56 <sup>d</sup>	7.24 ± 0.63 <sup>c</sup>	9.32 ± 0.24 <sup>b</sup>	14.88 ± 1.01 <sup>a</sup>
<i>A. indicum</i> leaf extract	<i>E. coli</i>	6.36 ± 0.82 <sup>d</sup>	10.24 ± 0.55 <sup>c</sup>	14.39 ± 0.79 <sup>b</sup>	17.42 ± 1.07 <sup>a</sup>
	<i>P. aeruginosa</i>	5.14 ± 0.47 <sup>d</sup>	7.43 ± 0.26 <sup>c</sup>	9.66 ± 0.36 <sup>b</sup>	13.15 ± 0.88 <sup>a</sup>
	<i>S. aureus</i>	2.07 ± 0.13 <sup>d</sup>	4.03 ± 0.13 <sup>c</sup>	5.96 ± 0.47 <sup>b</sup>	8.63 ± 0.57 <sup>a</sup>
	<i>B.subtilis</i>	3.98 ± 0.26 <sup>d</sup>	5.12 ± 0.67 <sup>c</sup>	7.06 ± 0.68 <sup>b</sup>	12.64 ± 0.69 <sup>a</sup>

Values mean ± SD indicates the replicates of three experiments.

\*Mean values within the column followed by the same ANOVA letter in superscript are significantly different at P < 0.05 level.

process. They constitute both primary and secondary antioxidants [42]. At higher concentration (120 µg/mL) of L-ascorbic acid displayed 97.15 ± 2.25 % superior reducing power activity flowed by Ai-FeO NPs 94.81 ± 2.70 %, which are summarized in Fig. 3B. The IC<sub>50</sub> values of reducing power activity for Ai-FeO NPs and L-ascorbic acid were determined from the plotted regression graph is found to be 56 µg/ml and 47 µg/ml, respectively (Table 1).

Nitric oxide (NO) is a bioregulatory molecule that is vital in the neurological, immunological, and cardiovascular systems. Fig. 3C revealed higher concentration (120 µg/mL) of BHT displayed 92.32 ± 1.74 % superior NO scavenging activity flowed by Ai-FeO NPs 89.92 ± 2.62 %. At lower concentration (20 µg/mL) of BHT displayed 12.63 ± 0.81 % superior NO scavenging activity flowed by Ai-FeO NPs 7.16 ±

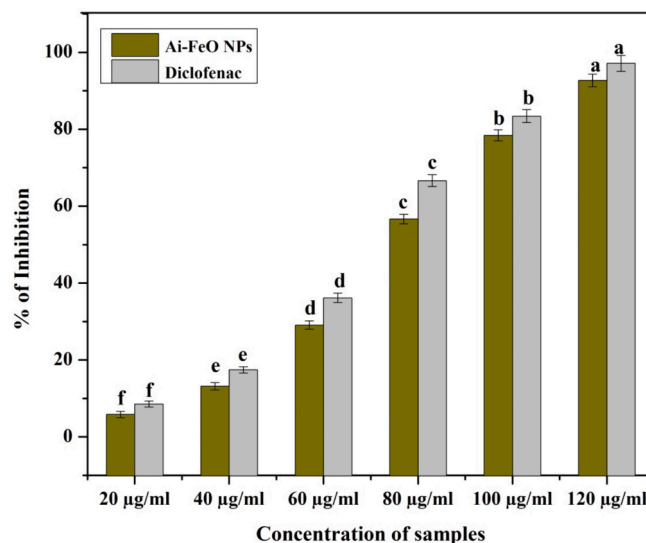


Fig. 5. (A) Anti inflammation of green synthesized Ai-FeO NPs and diclofenac against albumin, the data represents the mean values of three independent experiments and are presented as mean ± SD of the absorbance. Different letters indicated the significance differences (ANOVA followed by Tukey's HSD,  $\alpha = 0.005$ ).

Table 3

Cytotoxic activity of Cisplatin and Ai-FeO NPs (µg/ml).

S. No	Samples	MDA-MB-231(IC <sub>50</sub> )
1	Cisplatin	85.32 ± 2.72
2	Ai-FeO NPs	66.55 ± 1.46

IC<sub>50</sub> – Values of respective samples at 48 h.

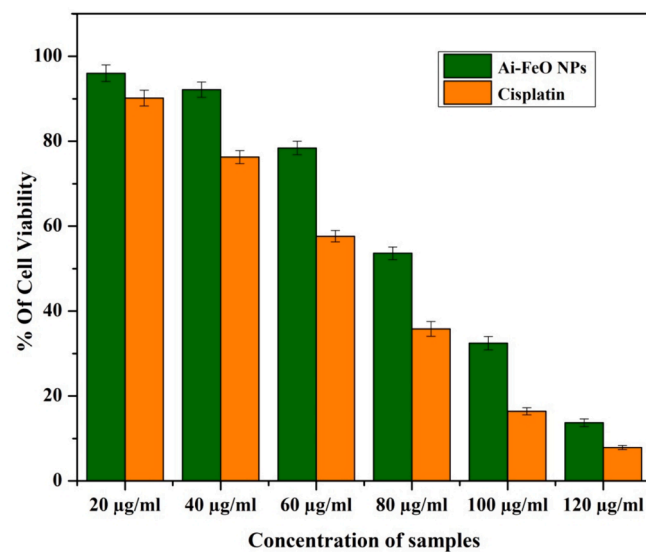
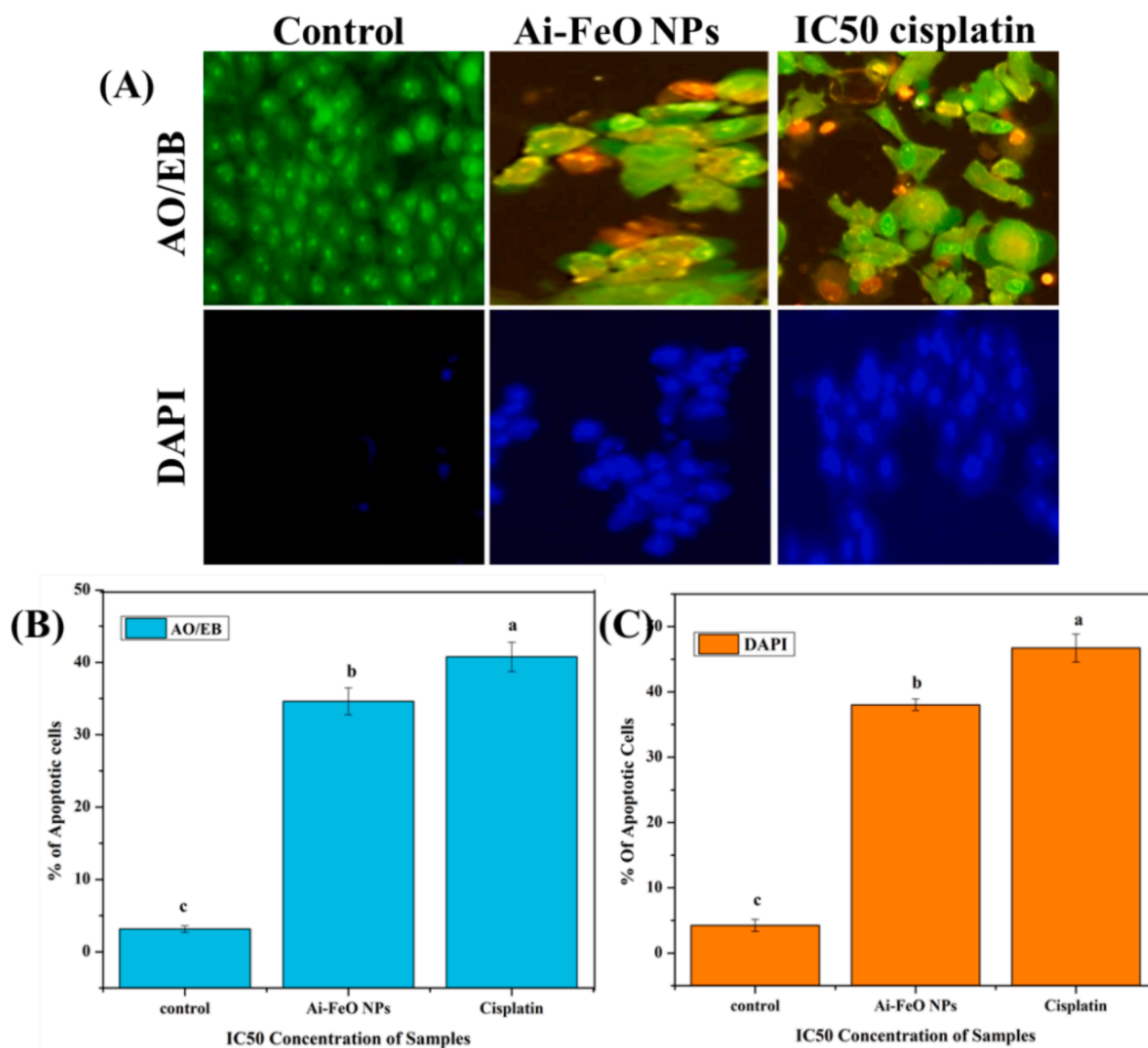


Fig. 6. Cytotoxic potential of *A.indicum* synthesized Ai-FeO NPs and Cisplatin against MDA-MB-231 cells. The data represents the mean values of three independent experiments and are presented as mean ± SD of the absorbance.

0.62 %. In addition, IC<sub>50</sub> values for NO scavenging activity for Ai-FeO NPs and BHT were determined from the plotted regression graph is found to be 75 µg/ml and 58 µg/ml, respectively as indicated in Table 1. Phytochemicals in the extract were responsible for the reducing power activity [50]. The results obtained in this study were similar to those of



**Fig. 7.** Bright field and fluorescence microscopy of MDA-MB-231 cells treated with IC<sub>50</sub> concentrations of Ai-FeO nanoparticles and cisplatin; (A) Fluorescence microscopy of AO/EtBr-stained control cells Ai-FeO NPs and Cisplatin treated cells (live cells in green color and dead cell in orange color, in the treated sample). DAPI nuclear staining of control cells and Ai-FeO NPs Cisplatin treated cells exhibited a condensed form of nuclear materials in apoptotic cells. (B, C) Histogram representation of the percentage of total observed apoptotic cells against the IC<sub>50</sub> concentration of Ai-FeO NPs and Cisplatin formulated on selected MDA-MB-231 cells after 48 h. The data represents the mean values of three independent experiments and are presented as mean  $\pm$  SD of the absorbance. Different letters indicated the significance differences (ANOVA followed by Tukey's HSD,  $\alpha = 0.005$ ).

Dipankar and Murugan [51] reported earlier.

#### 4.3. Antibacterial (zone of inhibition) activity of Ai-FeO NPs

The agar well diffusion method was used to evaluate the antibacterial activity of the synthesized Ai-FeO NPs and *A. indicum* leaf extract (Fig. 4). The zone of inhibition around the well containing different concentrations of samples was examined. The results show that the highest dose of Ai-FeO NPs was effective against both Gram-positive (*S. aureus*, *B. subtilis*) and Gram-negative bacteria (*E. coli*, *P. aeruginosa*), which are represented in Table 2. The data reported in Table 2 indicated that aqueous extract of *A. indicum* leaf extract had very low inhibitory effects on all the tested strains. The higher concentration of Ai-FeO NPs (100  $\mu\text{g}/\text{mL}$ ) displayed DIZs for *E. coli*, *P. aeruginosa*, *S. aureus*, and *B. subtilis* are  $19.25 \pm 1.08$  mm,  $16.56 \pm 0.6$  mm,  $12.63 \pm 1.07$  mm and  $14.88 \pm 1.01$  respectively, which are shown in Fig. 4. At lower concentration of Ai-FeO NPs showed DIZs for *E. coli*, *P. aeruginosa*,

and *S. aureus*, *B. subtilis* are  $8.12 \pm 0.72$  mm,  $5.48 \pm 0.67$  mm,  $3.51 \pm 0.72$  and  $4.68 \pm 0.56$ . In general, iron oxide nanoparticles shown antibacterial activity due to their ability of reactive oxygen species (ROS) production, cell membrane breakage and DNA damage, which leads to the cell death [23,52]. It was found that green synthesized FeO nanoparticles were more effective against Gram-negative bacteria than Gram-positive bacteria. Similarly, the green synthesized Ag doped CeO<sub>2</sub> showed enhanced antibacterial properties on *E. coli* and *S. aureus* [53]. The structural and compositional differences in the membranes of these two types of bacteria may lead to the difference in their activity [54,55]. These green synthesized FeO NPs would find it harder to permeate Gram-positive bacteria thicker peptidoglycan cell walls than Gram-negative bacteria, as there would be less of an antibacterial reaction [40].

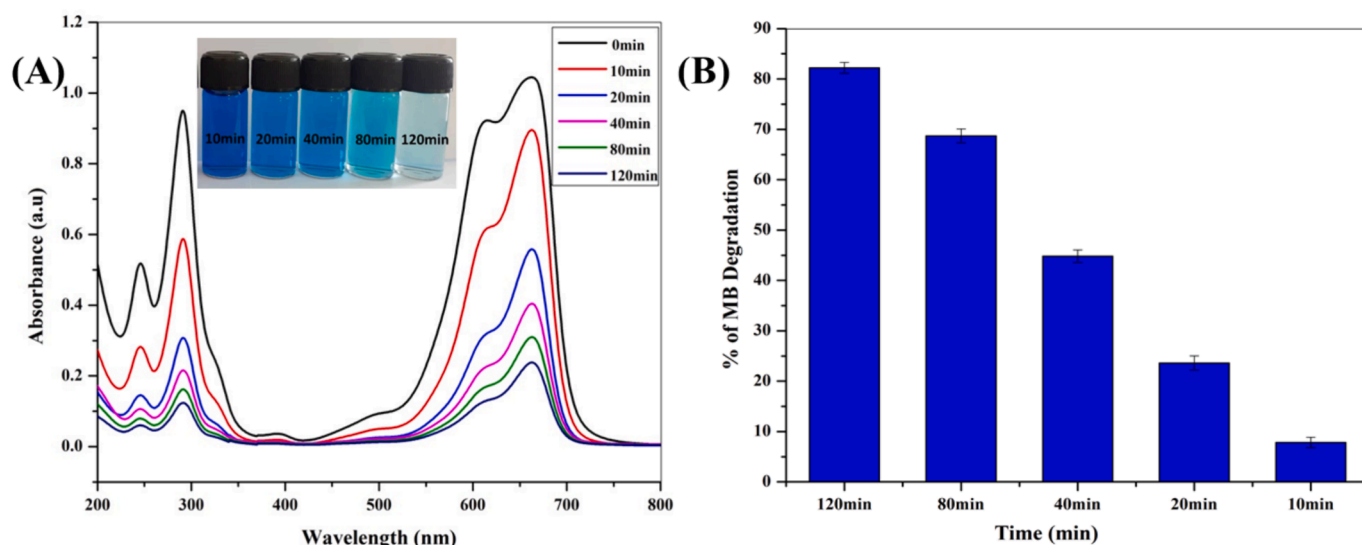


Fig. 8. Photocatalytic activity of *A.indicum* synthesized Ai-FeO nanoparticles.

#### 4.4. In-vitro anti-inflammatory activity

The immune system of the body automatically produces inflammation in reaction to a variety of pathogens, irritants, damaged cells, and damaging stimuli. Protein breakdown can also result in inflammation. In this work, we have evaluated the anti-inflammatory potential of Ai-FeO NPs using Albumin denaturation assays. Due to the external stress, proteins get denaturated and lose their biological functions as well as their secondary and tertiary structures. Fig. 5 revealed the green synthesized Ai-FeO NPs had a dose dependant anti-inflammatory activity. The activity of Ai-FeO NPs on Bovine Serum Albumin was increased and directly proportional to the concentration (20  $\mu\text{g}/\text{mL}$ –5.82 %; 40  $\mu\text{g}/\text{mL}$ –13.16 %; 60  $\mu\text{g}/\text{mL}$ –29.08 %, 80  $\mu\text{g}/\text{mL}$ –56.66, 100  $\mu\text{g}/\text{mL}$ –78.42 and 120  $\mu\text{g}/\text{mL}$ –92.71 %). Furthermore, diclofenac revealed a minimum inhibition of 8.54 % at a concentration of 20  $\mu\text{g}/\text{mL}$  and maximum inhibition of 97.15 % at a 120  $\mu\text{g}/\text{mL}$  concentration as shown in Fig. 5. Our results are well matched with Alahmdi et al. [19] revealed that zinc oxide nanoparticles from *Clitoria ternatea* flower extract exhibited significant anti-inflammatory activity in a dose dependent manner,

#### 4.5. Cytotoxic effect of Ai-FeO NPs using MDA-MB-231 cell line

The MTT assay is used to evaluate cellular metabolic activity which is the predictor of cell viability, proliferation, and cytotoxicity of a molecule. This colorimetric assay is principally based on the conversion of a yellow tetrazolium salt (three-four, five-dimethylthiazol-2-yl)-2, 5-diphenyl tetrazolium bromide or MTT) to red formazan crystals through metabolically active cells. The MTT is transformed to formazan by NADPH dependent oxidoreductase enzymes in live cells [56]. Fig. 6 demonstrates the different concentration of Ai-FeO nanoparticles in solutions at concentrations of 10  $\mu\text{g}/\text{ml}$ , 20  $\mu\text{g}/\text{ml}$ , 40  $\mu\text{g}/\text{ml}$ , 60  $\mu\text{g}/\text{ml}$ , 80  $\mu\text{g}/\text{ml}$  and 100  $\mu\text{g}/\text{ml}$ , giving viability reports of 96 %, 92 %, 78 %, 53 %, 32 % and 13 %. On the other hand, commercial drug in solutions at different concentrations like 10  $\mu\text{g}/\text{ml}$ , 20  $\mu\text{g}/\text{ml}$ , 40  $\mu\text{g}/\text{ml}$ , 60  $\mu\text{g}/\text{ml}$ , 80  $\mu\text{g}/\text{ml}$  and 100  $\mu\text{g}/\text{ml}$ , giving viability reports of 90 %, 76 %, 57 %, 35 %, 16 % and 7 %. From these results, we conclude that the green synthesized FeO NPs had a great cytotoxic effect against MDA-MB-231 cells, but slightly lower cytotoxic compared to commercial drug. The  $\text{IC}_{50}$  values of MDA-MB-231 in the presence of Ai-FeO NPs and cisplatin were calculated to be 85 and 66  $\mu\text{g}/\text{ml}$ , respectively (Table 3). Nanoparticles can pass across the cellular membrane and influence the mRNA expression of suppression genes, causing the production of reactive oxygen species (ROS) in cells to increase [57]. Unexpectedly, more

external ROS are produced by metal oxide nanoparticles, despite the fact that this is one of the signs of a severe cytotoxicity effect [58].

##### 4.5.1. Dual staining (AO/EB) and DAPI analysis of apoptosis

The ability of Ai-FeO NPs ( $\text{IC}_{50}$ ) to destabilise cell membranes was assessed by using the AO/EB assay in comparison with the controls. The results showed that the early apoptosis was increased, and number of viable cells decreased after the treatment. AO and EB dual staining was employed to search for various apoptotic indicators in nucleate alterations (Fig. 7a). The percentage of cellular damage was reported to be  $40.76 \pm 2.03$  and  $34.62 \pm 1.85$ , in treated and control cells respectively. Apoptosis-induced early cell shrinkages, DNA fragmentation, perinuclear condensed chromatin, and late apoptosis-induced fragmented chromatin and cell nuclei disintegration were clearly visible in green synthesized Ai-FeO NPs treated cells. The DAPI stained negative control and Ai-FeO NPs treated cells (Conc. same as  $\text{IC}_{50}$  value) and positive control (cisplatin) treated MDA-MB-231 were also analysed there was a significant number of apoptotic nuclei in Ai-FeO NPs treated cells (Fig. 7b). The nuclear condensation rate due to Apoptosis was found to be  $38.03 \pm 1.08$  and  $46.72 \pm 2.14$  respectively (figure c).

#### 4.6. Photocatalytic activity of Ai-FeO NPs

The pharmaceutical, textile, and dyeing sectors essentially employ a lethal synthetic colour known as methylene blue. Methylene blue has a potential to upset the balance of aquatic and biological systems [59]. In this work, we have carried out the photocatalytic activity of green synthesized Ai-FeO NPs against methylene blue (MB). The following conditions were used to study the degradation of MB: The reaction mixture of 50 ml of MB and 25 mg of Ai-FeO nanoparticles were exposed to sunlight for various time intervals. The MB was gradually photo degraded during this process, and the colour of the solution changed from an initial deep blue to nearly translucent, the absorbance intensity gradually diminishes with longer contact durations, that indicates the photocatalytic degradation (Fig. 8). In case of control where Ai-FeO NPs was absent methylene blue could not degrade under the irradiation of sun light. The percentage of MB that was degraded at different time intervals (120 min, 80 min, 40 min, 20 min, and 10 min) and their percentages of removal 82.24, 68.42, 44.68, 23.62, and 7.83 %, respectively are shown in Fig. 6b. Similarly, *Gazania rigens* synthesized Cu-Ni bimetallic nanoparticles efficiently degrade the methylene blue under UV light irradiation after 19 mins treatment with a rate constant of  $0.2505 \text{ min}^{-1}$  [60]. Our result indicated that the presence of green



synthesised Ai-FeO NPs was causing the methylene blue degradation in time-dependent manner.

## 5. Conclusion

Recently, plant extracts have been employed in the development of environmentally friendly nanoparticles via green synthesis approach for several therapeutic purposes as they possess low toxicity and are cost-effective. Nowadays, nanoparticles are used in many aspects of medicinal research such as antimicrobial, cancer research, drug delivery etc. Keeping this in mind we focused our research to green synthesize FeO nanoparticles using leaf extracts of *A. indicum*. Further, the physical and chemical properties of obtained Ai-FeO NPs were thoroughly investigated by UV–vis spectroscopy, XRD, FTIR, SEM, HR-TEM, and EDAX analysis. Furthermore, our results demonstrated that the green synthesized Ai-FeO NPs using *A. indicum* displayed appreciable biomedical properties, including antimicrobial, antioxidant, anti-inflammatory, and anti-apoptotic activities. Overall, according to our study the *A. indicum*-synthesized Ai-FeO NPs can be a potential candidate for variety of therapeutic drugs especially as biosafe and cost-effective antioxidant, antibacterial, and a significant anti-breast cancer drug.

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## CRediT authorship contribution statement

**Chellasamy Panneerselvam:** . **Mohammed Ali Alshehri:** Data curation. **Ahmed Saif:** . **Uzma Faridi:** Writing – original draft, Methodology. **Syed Khasim:** Writing – original draft, Methodology. **Zuhair M Mohammedsalem:** Data curation. **Humaira Parveen:** Writing – original draft. **Noha Omer:** Writing – review & editing, Data curation. **Abdulrahman Alasmari:** Data curation. **Sayed Mukhtar:** Writing – review & editing, Writing – original draft. **Hatem A. Al-Aoh:** Conceptualization.

## Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## Data availability

Data will be made available on request.

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