

# Antibacterial Activity of Synergistic Effect of colicin and Gold Nanoparticles against *Klebsiella pneumonia*

Zaid shaker Al-mawlawi<sup>1</sup>, Hind Hussein Obaid<sup>1</sup>

<sup>1</sup>Department of Biology, College of Science, University of Baghdad, Baghdad, Iraq

## ABSTRACT

Fifty of urine samples were collected from patients with urinary tract infection (UTI). The samples were collected from AL- Yarmuk hospital in Baghdad. All of the isolates were diagnosed using biochemical test and vitek. The result showed that 30 (60%) isolates identified as *E.coli* from 50 urine samples. The colicinogenic isolates were determined using cup assay methods. The results showed that 10 out of 30 isolates (33.3%) were detect as colicin producers from 30 isolate identified as *E.coli* depending on the clear zone that observed against the sensitive isolate. Colicin was extracted from the efficient isolate. Colicin activity (320 U/ml) was determined by well assay method. The protein concentration (520 µg/ml) estimated by using Bradford assay. The watery extract of Chilli papers (*Capsicum baccatum*) was extracted and used it as reducing and capping agent for gold nanoparticles synthesis. The characterization of the gold nanoparticles was done by UV-Visible Spectrophotometer, Transmission Electron Microscope (TEM), and resulting spherical nanoparticles with diameter ranging between (35-70 nm). The antibacterial activity of colicin alone and gold nanoparticles alone and combination of colicin with gold nanoparticles against ten isolates *Klebsiella pneumonia* isolated from urine samples, using tube method.

**Keywords:** Colicin, gold nanoparticles, Chilli papers, antibacterial

## INTRODUCTION

*Escherichia coli* is an important genus belong to the family Enterobacteriaceae that found in human and animal intestine because its share in facilitation of digestion and fermentation of food<sup>1-2</sup>. There are some bacteria secrete substances which as a protein in nature used as a defensive mechanism against related or another genus of bacteria, from these the colicin that secrete by *Escherichia coli* and these character benefit in use the colicin as antibiotic in treated some diseases and inhibition growth of some bacteria<sup>3</sup>. Some researcher define the colicin as a protein substance that secreted by different genus of bacteria and characterized by have bactericidal activity against other strains, and mode of action depend on specific receptor in sensitive cell

for these colicin<sup>4,5</sup>. There are some scientist said that the colicin it's a toxic protein produce by some strain of *E .coli* and became active against related or nearby strain<sup>6</sup>. The colicin is a weapon that the *E .coli* uses it in competitive war against other *E .coli* or other bacteria to get nutrient<sup>7</sup>. Colicins are proteins that consist from three specific domains, amino-terminal translocation (T) domain, central receptor-binding (R) domain, and carboxy-terminal cytotoxic (C) domain<sup>8,9</sup>. The colicin particles was proteins in nature, also some colicins are composed from protein with carbohydrate or lipids but its few, for these we find the colicin particle is similar to any protein particle composed from amino acids that form small peptide chain and these peptide bind with each other to produce three dimensional shape of protein<sup>10</sup>. The colicin made and secreted in few amount in the bacterial cell that have plasmid only<sup>11</sup>, but various external factors affecting the regulatory expression of colicin have been revised extensively by Cascales *et al.*, in 2007<sup>8</sup>. The term of nanoparticles (NPs) usually gain for the particles that have size ranging from 1-100 nanometers (nm). The raw metals have inert properties and when decrease in the sizes of the metals to the

### Corresponding author:

**Hind Hussein Obaid.**

Department of Biology, College of Science, University of Baghdad, Baghdad, Iraq;

E-mail: hind\_3000@yahoo.com.

atomic level their properties changing to the benefit form<sup>12</sup>, the nanoparticles have unique physico-chemical and biological properties which can be used in different suitably applications<sup>13</sup>. Certain nanopowders possess antimicrobial properties. When these powders contact cells of *E. coli*, or other bacteria species and viruses, over 90% are killed within a few minutes<sup>14</sup>. Due to their antimicrobial effect, nanoparticle of silver and titanium dioxide (<100nm) are assessed as coatings for surgical masks. Zinc Oxide nano particles can decrease the antibiotic resistance and enhance the antibacterial activity of Ciprofloxacin against microorganism, by interfering with various proteins that are interacting in the antibiotic resistance or pharmacologic mechanisms of drugs<sup>15</sup>. The application of using the gold nanoparticles in biomedical products is being developed for drug delivery, cancer therapy, diagnostic devices, biosensing, and bacterial growth inhibition<sup>16</sup>.

## MATERIALS AND METHOD

### Bacterial isolation and identification

Fifty of urine samples were collected from patients with urinary tract infection (UTI) the samples were collected from AL- Yarmuk hospital in Baghdad. All urine Samples were collected in sterilized containers, in the laboratory within aseptic conditions; the collected samples were streaked directly on MacConkey agar and EMB agar (Himedia/India) and incubated for 24h at 37°C. Pink colonies were picked. Further identification tests included the morphological characteristics and biochemical tests were carried out depending on Harley and Prescott in 2002, and Brenner in 2005<sup>17,18</sup>.

### Extraction of crude non-bound colicin<sup>19</sup>.

After determination of colicinogenic *E. coli* using cup assay methods [20] the colicin extracted from efficient isolate as following:

The overnight culture of bacterial isolates in volume 2.5 ml of nutrient broth was used to inoculate 100 ml of sterile nutrient broth supplemented with 5 % glycerol and incubated in shaker incubator for 14 hrs at 37 C.

At cell density of about  $3 \times 10^8$  cells/ ml (14 hrs. incubation of late log phase), Mitomycin- C was added at a final concentration of 2 µg / ml, and incubate in shaker incubator for another 3 hrs.

The culture was centrifuged at 5000 rpm for 30 min

in cooling centrifuge. The supernatant was taken for assay of colicin using well methods<sup>21</sup>, and the concentration of protein was determined<sup>22</sup>.

Chloroform ((10 %) was added for killing any cells may be found in the supernatant. All supernatants were cultured on Brain heart infusion agar in order to confirm the absence of *E. coli* cells.

### Synthesis and characterization of Gold Nanoparticles (Au NPs).

Gold nanoparticles were synthesis by green method using chilli papers (*Capsicum baccatum*) as reducing and stabilizing agent<sup>23</sup>. The morphological feature of gold nanoparticles identified using UV-Vis Spectral Analysis [24] and Transmission Electron Microscope (TEM) [25].

### Antibacterial activity

Antibacterial activity of colicin alone and gold nanoparticles alone and combination of (colicin + gold nanoparticles) were investigated by using an tubes method<sup>26</sup> against ten isolates of *Klebsiella pneumonia* that isolated from burn samples.

## RESULTS AND DISCUSSION

### Isolation and identification:

Thirty isolates (60%) identified as *E. coli* from 50 urine sample and others were not *E. coli*. The highest percentage of *E. coli* isolation from UTI revealed that *E. coli* was the main causative agent of UTI. the *E. coli* cause 90% of the urinary tract infection<sup>27</sup>.

### Determination efficient colicinogenic *E. coli*.

The main purpose of collection and identification of *E. coli* was for determination of the efficient isolate that able to produce colicin. There are several methods can be used for screening about colicin but in this study we used cup assay method and resulting 10 out of 30 isolates identified as *E. coli* isolate (33.3%) as colicin producers according to inhibition zone resulting from these processes one efficient isolate had been selected from ten colicin producers isolates, because it gave a higher inhibition zone (23 mm) against the sensitive isolate among producers isolate.

Characterization of Synthesis Gold Nanoparticles (Au NPs).

The first method for characterization of biosynthesis gold nanoparticles was UV–Vis spectrophotometry. The figure (1) explained all obtained results. Three UV–Vis tests were piloted in different time intervals and observed that the color changed of the gold nanoparticles with a time progresses. In the first (30) minutes no color change with no peak observed; After four hours the color change from yellow to red with a peak showed in wave length at (552.50 nm) and absorption (0.694), after 24 hours the color converted to a ruby red and the peak detected in wave length at(550.00 nm) and absorption value (1.490). The result showed revealed that the color change play an important role in detected the formation of nanoparticles, and this was confirmed by the appearance of the peaks in conjunction with the absorbance during the time progresses. The peaks were appears gave a spectroscopic signature to form a surface plasmon resonance (SPR) of gold nanoparticle <sup>23</sup>. The peaks shifted not much with time from (552.50nm) to (550.00 nm) with increase in absorbance from (0.694) to (1.490) were revealed a linked point between the more reduction reaction and formation nanoparticles[28]. This study was agreement with study reported by Kumar *et al.*, in 2015 that when appearance the dark color confirm the formation of nanoparticles and efficient reduction the to.

**Determination of Antibacterial activity of colicin, gold nanoparticles, combination of (colicin + gold nanoparticles) against *Klebsiella pneumonia* isolated from urine samples, using tube method**

The activity of colicin against 10 isolates of *Klebsiella pneumonia* is shown in table (1), and showed the different concentration of colicin has antibacterial activity for different strains of *Klebsiella pneumonia*. The highest colicin activity appears at concentration (16.25 µg/ ml) with activity 32U/ml against two strains (K1 and K3). The recent work revealed the crude extract of colicin extraction from producer *E. coli* showed a wide activity spectrum on other gram negative bacteria in different concentrations, this was because the colicin active against related or nearby strain. The table (2) showed the affected of *Klebsiella pneumonia* by gold nanoparticles, and you will see the higher concentration of gold nanoparticles(1395 ng/ ml) inhibit growth all strains, and the higher activity of gold nanoparticles was (1024 U/ml) at concentration (2.73 ng/ ml) as in isolates (K1 and K2). The table (3) represents the activity of the combination against *Klebsiella pneumoniae* and shows that most of the isolates affected by different concentration of combination.

**Table 1. Antimicrobial effect of crude colicin extracted from *E. coli* against *Klebsiella pneumonia* isolated from urine samples, using tube method.**

Isolates		K1	K2	K3	K4	K5	K6	K7	K8	K9	K10
Dilution	Protein conc. µg/ ml	(-) Growth / (+) No growth									
1/2	260	+	+	+	+	+	+	+	+	+	+
1/4	130	+	-	+	+	+	+	+	+	+	+
1/8	65	+	-	+	+	+	+	+	-	-	+
1/16	32.5	+	-	+	+	+	+	-	-	-	-
1/32	16.25	+	-	+	-	-	-	-	-	-	-
1/64 to 1/1024	8.125 to 0.507	-	-	-	-	-	-	-	-	-	-
Activity U/ml		32	2	32	16	16	16	8	4	4	8

**Table 2. Antimicrobial effect of Gold Nanoparticles Against *Klebsiella pneumonia* isolated from urine samples, using tube method.**

Isolates		K1	K2	K3	K4	K5	K6	K7	K8	K9	K10
Dilution	GoldNanoparticles conc. ng/ ml	(-) Growth / (+) No growth									
1/2	1395	+	+	+	+	+	+	+	+	+	+
1/4	697.5	+	-	+	+	+	+	+	+	+	+
1/8	348.75	+	-	-	-	-	-	-	-	-	+
1/16	174.38	+	-	-	-	-	-	-	-	-	+
1/32	87.19	+	-	-	-	-	-	-	-	-	+
1/64	43.59	+	-	-	-	-	-	-	-	-	+
1/128	21.81	+	-	-	-	-	-	-	-	-	+
1/256	10.91	+	-	-	-	-	-	-	-	-	+
1/512	5.45	+	-	-	-	-	-	-	-	-	+
1/1024	2.73	+	-	-	-	-	-	-	-	-	+
1/2048 to 1/4048	1.36 to 0.68	-	-	-	-	-	-	-	-	-	-
Activity U/ml		1024	2	4	4	4	4	4	4	4	1024

**Table 3. Synergistic effect of colicin and Gold Nanoparticles against *Klebsiella pneumoniae* isolated from urine samples, using tube method.**

Isolates			K1	K2	K3	K4	K5	K6	K7	K8	K9	K10
Dilution	Protein conc. µg/ ml	GoldNanoparticles conc. ng/ ml	(-) Growth / (+) No growth									
1/4	130	697.5	+	+	+	+	+	+	+	+	+	+
1/8	65	348.75	+	-	+	+	+	+	+	+	+	+
1/16	32.5	174.38	+	-	+	+	+	+	+	+	+	+
1/32	16.25	87.19	+	-	+	+	+	+	+	+	+	+
1/64	8.125	43.59	+	-	+	+	+	+	-	+	+	+
1/128	4.06	21.81	+	-	+	-	+	-	-	+	+	-
1/256	2.03	10.91	+	-	+	-	+	-	-	-	+	-
1/512	1.015	5.45	+	-	-	-	-	-	-	-	+	-
1/1024	0.507	2.73	+	-	-	-	-	-	-	-	+	-
1/2048	0.253	1.36	+	-	-	-	-	-	-	-	-	-
1/4096 to 1/8192	0.126 to 0.063	0.68 to 0.34	-	-	-	-	-	-	-	-	-	-
Activity U/ml			2048	4	256	64	256	64	32	128	1024	64

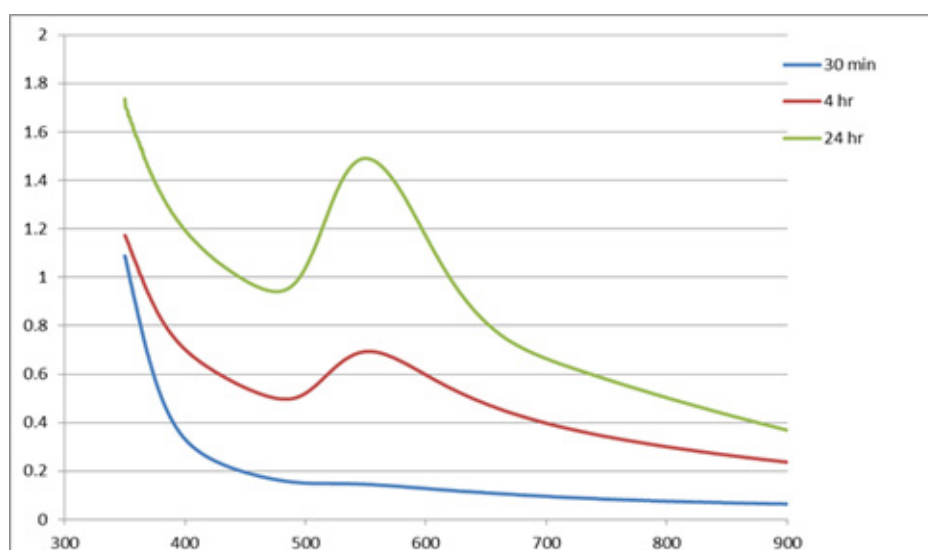


Figure 1. UV-Vis spectrophotometry of gold nanoparticles synthesis by chilli papers extract.

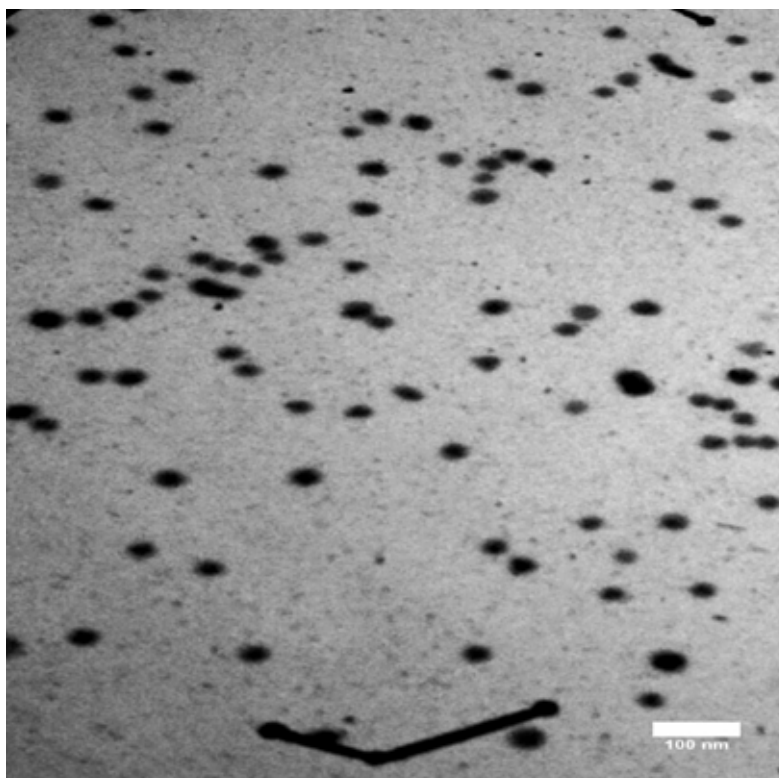


Figure 2. Transmission electron microscopic (TEM) images of Au NPs Synthesized using chilli papers extract.

## CONCLUSION

The characterization of the gold nanoparticles was done by UV-Visible Spectrophotometer, Transmission Electron Microscope (TEM), and resulting spherical nanoparticles with diameter ranging between (35-70 nm). The antibacterial activity of colicin alone and gold nanoparticles alone and combination of colicin with gold nanoparticles against ten isolates *Klebsiella pneumonia* isolated from urine samples, using tube method.

**Financial Disclosure:** There is no financial disclosure.

**Conflict of Interest:** None to declare.

**Ethical Clearance:** All experimental protocols were approved under the Department of Biology, College of Science, University of Baghdad, Iraq and all experiments were carried out in accordance with approved guidelines.



## REFERENCES

1. Feng P, Weagant S, Grant M. Enumeration of *Escherichia coli* and the Coliform Bacteria. *Bacteriological Analytical Manual* (8th Ed.). FDA/Center for Food Safety and applied nutrition. 2002.
2. Mainil J. *Escherichia coli* virulence factors. *Veterinary immunology and immunopathology*. 2013; 152(1): 2-12.
3. Yamazaki K, Suzuki M, Kawai Y, Inoue N. Purification and characterization of a novel class IIa bacteriocin, piscicocin CS526, from surimi-associated *Carnobacterium piscicola* CS526. *Applied and environmental microbiology*. 2005; 71(1): 554-557.
4. Pugsley AP, Schwartz M. Colicin E2 release: lysis, leakage or secretion? Possible role of a phospholipase. *The EMBO journal*. 1984; 3(10): 2393.
5. Cavard DA, Lazdunski C. Colicin cleavage by OmpT protease during both entry into and release from *Escherichia coli* cells. *Journal of bacteriology*. 1990; 172(2): 648-652.
6. Luirink JO, Clark DM, Ras J, Verschoor EJ. pCloDF13-encoded bacteriocin release proteins with shortened carboxyl-terminal segments are lipid modified and processed and function in release of cloacin DF13 and apparent host cell lysis. *Journal of bacteriology*. 1989; 171(5): 2673-2679.
7. Kirkup BC, Riley MA. Antibiotic-mediated antagonism leads to a bacterial game of rock-paper-scissors in vivo. *Nature*. 2004; 428(6981): 412-414.
8. Cascales E, Buchanan SK, Duché, D. Colicin biology. *Microbiology and Molecular Biology Reviews*. 2007; 71(1): 158-229.
9. Kleanthous C. Swimming against the tide: progress and challenges in our understanding of colicin translocation. *Nature Reviews Microbiology*. 2010; 8(12): 843-848.
10. Fath MJ, Zhang LH, Rush J, Kolter R. Purification and characterization of colicin V from *Escherichia coli* culture supernatants. *Biochemistry*. 1994; 33(22): 6911-6917.
11. Pugsley AP, Schwartz M. A genetic approach to the study of mitomycin-induced lysis of *Escherichia coli* K-12 strains which produce colicin E2. *Molecular and General Genetics* MGG. 1983; 190(3): 366-372.
12. El-Sayed IH, Huang X, El-Sayed MA. Selective laser photo-thermal therapy of epithelial carcinoma using anti-EGFR antibody conjugated gold nanoparticles. *Cancer letters*. 2006; 239(1): 129-135.
13. Sanvicens, N., & Marco, M. P. 2008. Multifunctional nanoparticles—properties and prospects for their use in human medicine. *Trends in biotechnology*. 26(8): 425-433.
14. Bahadori M, Mohammadi F. Nanomedicine for respiratory diseases. *Tanaffos*. 2012; 11(4): 18.
15. Banoee M, Seif S, Nazari ZE. ZnO nanoparticles enhanced antibacterial activity of ciprofloxacin against *Staphylococcus aureus* and *Escherichia coli*. *Journal of Biomedical Materials Research Part B: Applied Biomaterials*. 2010; 93(2): 557-561.
16. De Jong W. H., & Borm, P. J. 2008. Drug delivery and nanoparticles: applications and hazards. *International journal of nanomedicine*, 3(2), 133.
17. Harley J, Prescott H. *Laboratory Exercises in Microbiology*, fifth Edition. The McGraw-Hill Companies. 2002.
18. Brenner DJ, Krieg NR, Staley JT. (Eds.) *Bergey's Manual of Systematic Bacteriology*, Volume Two: The Proteobacteria. Part B: The Gammaproteobacteria. second Edition. Springer Science+ Business Media, LLC, 233 Spring Street, New York, NY 10013, USA. 2005.
19. Herschman HR, Helinski DR. Purification and characterization of colicin E2 and colicin E3. *Journal of Biological Chemistry*. 1967; 242(22): 5360-5368.
20. Al-Qassab AO, Al-Khafaji ZM. Effect of different conditions on inhibition activity of enteric lactobacilli against diarrhea-causing enteric bacteria. *J. Agri. Sci*. 1992; 3(1): 18-26.
21. Smajs D, Pilsel H. Colicin U, A novel Colicin Produced by *Shigella boydii*. *J. of Bacteriol*. 1997; 179: 4919- 28.
22. Bradford MM. A dye binding assay for protein. *Anal. Biochem*. 1976; 72: 248-254.
23. Kumar B, Smita K, Cumbal L. Phytosynthesis of gold nanoparticles using Andean Aji' (*Capsicum baccatum* L.). *Cogent Chemistry*. 2015; 1(1):

1120982‡

24. Haiss W, Thanh NT, Aveyard J. Determination of size and concentration of gold nanoparticles from UV– Vis spectra. *Analytical chemistry*. 2007; 79(11): 4215-4221‡
25. Reimer L. *Transmission electron microscopy: physics of image formation and microanalysis*. 2013.
26. Saxena G, Farmer S, Hancock RE, Towers GH. Antimicrobial compounds from *Alnus rubra*. *International journal of pharmacognosy*. 1995; 33(1): 33-36‡
27. Stapleton A. Novel mechanism of P-fimbriated *Escherichia coli* virulence in pyelonephritis. *Journal of the American Society of Nephrology*. 2005; 16(12): 3458-3460‡